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**United States Patent**  
**Chaudhuri , et al.****10,350,280**  
**July 16, 2019**

Methods to analyze genetic alterations in cancer to identify therapeutic peptide vaccines and kits therefore

**Abstract**

The invention describes a method for identifying T-cell activating neo-epitopes from all genetically altered proteins. The mutated proteins contribute to neo-epitopes after they are proteolytically degraded within antigen presenting cells, such as dendritic cells and macrophages.

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*Parent Case Text*

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CROSS REFERENCE TO RELATED APPLICATIONS

The subject application claims priority under 35 U.S.C. 111(a) to PCT Application No. PCT/US2017/049773, filed Aug. 31, 2017, which claims the benefit of U.S. Provisional Application No. 62/382,179 filed Aug. 31, 2016, all of which are incorporated herein by reference in their entireties into the present patent application for all purposes.

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*Claims*

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What is claimed is:

1. A method of selecting mammalian tumor immunogenic peptides from genetically altered proteins expressed by a mammalian tumor cell or a mammalian tumor tissue from a subject, which method comprises: a) obtaining a sample from the subject; b) identifying genetically altered proteins expressed by a mammalian tumor cell or a mammalian tumor tissue in the sample through nucleic acid sequences encoding the altered proteins; c) producing peptide fragments comprising at least one amino acid mutation from the genetically altered proteins so identified in step (b), so as to obtain peptide variants associated with the mammalian tumor cell or the mammalian tumor tissue; d) selecting peptide variants from step c which bind T-cell receptor (TCR) comprising: i) selecting peptide variants with a pre-defined length of 9 amino acids; ii) characterizing the peptide variants in silico by selecting and matching features associated with an amino acid

at each position of the peptide variants with selected pre-defined features of hydrophobic and helix/turn motif for each position of peptides recognized by TCR associated with CD8+ T-cell, so as to obtain predictive ability of the peptide variants to interact with the TCR and identifying peptide variants having the matching selected pre-defined features; and iii) selecting the peptide variants in step d.ii by: A. applying RICJ880105 and QIAN880107 helix/turn motif scales and a YUTK870103 hydrophobic scale and selecting peptide variants having: (a) at positions 5 and 6 an average value greater than 0.5 using a RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn scale, and (c) at positions 8 and 9 an average value less than or equal to 17.75 using a YUTK870103 hydrophobic scale; B. applying RICJ880105 and QIAN880107 helix/turn motif scales and a YUTK870103 hydrophobic scale and selecting peptide variants having: (a) at positions 5 and 6 an average value greater than 0.5 using a RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value less than or equal to -0.34 using a FNSA.2 charge of side chain scale, and (e) at positions 6 and 7 an average value less than or equal to 0.2055 using a VASM830101 scale for spatial flexibility of side chain and spatial flexibility of main chain; C. applying using RICJ880105 and QIAN880107 helix/turn motif scales and a YUTK870103 hydrophobic scale and selecting peptide variants having: (a) at positions 5 and 6 an average value greater than 0.5 using a RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average greater than -0.77 using a QIAN880107 helix/turn motif scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value greater than -0.34 using a FNSA.2 charge of side chain scale, and (e) at positions 6 and 7 an average value less than or equal to -5.5 using a ROBB760108 helix/turn scale; D. applying RICJ880105 and QIAN880107 helix/turn motif scales and a YUTK870103 hydrophobic scale and selecting peptide variants having: (a) at positions 5 and 6 an average value greater than 0.5 using a RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn motif scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value greater than -0.34 using a FNSA.2 charge of side chain scale, (e) at positions 6 and 7 an average value greater than -5.5 using a ROBB760108 helix/turn scale, (f) at positions 1-9 an average value less than or equal to 45.56 using a NAKH920106 helix/turn motif scale, and (g) at positions 2 and 3 an average value greater than -0.055 using a QIAN880139 helix/turn motif scale; E. applying RICJ880105 and QIAN880107 helix/turn motif scales and a YUTK870103 hydrophobic scale and selecting peptide variants having: (a) at positions 5 and 6 an average value greater than 0.65 using a RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn motif scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value greater than -0.34 using a FNSA.2 charge of side chain scale, (e) at positions 6 and 7 an average value greater than -5.5 using a ROBB760108 helix/turn scale, (f) at positions 1-9 an average value greater than 45.56 using a NAKH920106 helix/turn motif scale, (g) at positions 2 and 3 an average value greater than -0.055 using a QIAN880139 helix/turn motif scale, (h) at positions 7 and 8 an average value less than or equal to -0.23 using a QIAN880138 helix/turn motif scale, and (i) at positions 1-9 an average value greater than 7.0 using a CHAM830103 steric scale; F. applying RICJ880105, QIAN880107, ROBB760108, NAKH920106 and QIAN880138 helix/turn motif scales and a YUTK870103 hydrophobic scale and selecting peptide variants having: (a) at positions 5 and 6 an average value greater than 0.5 using a RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn motif scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value greater than -0.34 using a FNSA.2 charge of side chain scale, (e) at positions 6 and 7 an average value greater than -5.5 using a ROBB760108 helix/turn scale, (f) at positions 1-9 an average value greater than 45.56 using a NAKH920106 helix/turn motif scale, (g) at positions 7 and 8 an average value greater than -0.23 using a QIAN880138 helix/turn motif scale, (h) at positions 1 and 2 an average value less than or equal to 0.625 using a MITS020101 solvent accessibility of an amino acid scale, (i) at positions 2 a value less than or equal to 0.144401 using PNSA.1.AUTO charge of side chain scale, (j) at position 2 a value greater than -0.303435 using a PNSA.1.AUTO charge of side chain scale, (k) at position 3 a value less than or equal to 6.8 using a KARS160118 amino frequency scale, and (l) at positions 8 and 9 an average value less than or equal to 18.04 using a YUTK870104 hydrophobic scale; G. applying RICJ880105, QIAN880107, ROBB760108, NAKH920106 and QIAN880138 helix/turn motif scales and a YUTK870103 hydrophobic scale and selecting peptide variants having: (a) at positions 5 and 6 an average value greater than 0.5 using a RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn motif scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value greater than -0.34 using a FNSA.2

charge of side chain scale, (e) at positions 6 and 7 an average value greater than -5.5 using a ROBB760108 helix/turn scale, (f) at positions 1-9 an average value greater than 45.56 using a NAKH920106 helix/turn motif scale, (g) at positions 7 and 8 an average value greater than -0.23 using a QIAN880138 helix/turn motif scale, (h) at positions 1 and 2 an average value less than or equal to 0.625 using a MITS020101 solvent accessibility of an amino acid scale, (i) at position 2 a value less than or equal to 0.144401 using a PNSA.1.AUTO charge of side chain scale, (j) at position 3 a value greater than 6.8 using a KARS160118 amino frequency scale, and (k) at positions 5 and 6 an average value less than or equal to 17.92 using a YUTK870103 hydrophobic scale; or H. applying RICJ880105, QIAN880107, ROBB760108, NAKH920106 and QIAN880138 helix/turn motif scales and a YUTK870103 hydrophobic scale and selecting peptide variants having: (a) at positions 5 and 6 an average value greater than 0.5 using a RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn motif scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value greater than -0.34 using a FNSA.2 charge of side chain scale, (e) at positions 6 and 7 an average value greater than -5.5 using a ROBB760108 helix/turn scale, (f) at positions 1-9 an average value greater than 45.56 using a NAKH920106 helix/turn motif scale, (g) at positions 7 and 8 an average value greater than -0.23 using a QIAN880138 helix/turn motif scale, (h) at positions 1 and 2 an average value less than or equal to 0.625 using a MITS020101 solvent accessibility of an amino acid scale, and (i) at position 2 a value greater than 0.144401 using PNSA.1.AUTO charge of side chain scale; thereby, selecting mammalian tumor immunogenic peptides from genetically altered proteins expressed by the mammalian tumor cell or the mammalian tumor tissue.

2. The method of claim 1, wherein the immunogenic peptides are further selected by potential or ability to be produced inside the cell by processes comprising: a) determining the action of proteases, which are part of the proteasomal or immunoproteasomal complexes, based on the probability that processing event of the altered proteins will produce the immunogenic peptides so selected; and b) determining the entry of the immunogenic peptides into the endoplasmic reticulum compartment by binding to peptide transporters expressed on the surface of the compartment.

3. The method of claim 1, wherein in step (b) identifying the genetically altered proteins expressed by the mammalian tumor cell or the mammalian tumor tissue through nucleic acid sequences encoding the altered proteins comprises: a) identifying tumor variants from transcriptome analysis of the mammalian tumor cell or mammalian tumor tissue corresponding to protein coding and protein non-coding sequences; and b) performing conceptual translation or in silico translation of the coding sequences in step (a) so as to identify the genetically altered proteins expressed by the mammalian tumor cell or the mammalian tumor tissue.

4. The method of claim 3, wherein in step (a) identifying tumor variants from transcriptome analysis of the mammalian tumor cell or mammalian tumor tissue comprises a) determining nucleotide sequence of transcripts produced by the mammalian tumor cell or mammalian tumor tissue; and b) comparing the determined nucleotide sequence of transcripts in (a) with a reference nucleotide sequence of transcripts produced by mammalian non-tumor cell or mammalian non-tumor tissue, so as to identify nucleotide sequence changes in the protein coding and protein non-coding sequences; thereby, identifying tumor variants from transcriptome analysis of the mammalian tumor cell or mammalian tumor tissue.

5. The method of claim 3, further comprising performing genomic analysis for tumor variants in the sequence of the genome present in the mammalian tumor cell or the mammalian tumor tissue but absent or deficient in the mammalian non-tumor cell or the mammalian non-tumor tissue.

6. The method of claim 5, wherein the genomic analysis for tumor variants comprises determining nucleotide sequence of the genome or exome.

7. The method of claim 1, wherein in step (c) producing peptide fragments comprising at least one amino acid mutation from each genetically altered protein, so as to obtain peptide variants associated with the mammalian tumor cell or the mammalian tumor tissue comprises: a) defining length of the peptide fragments to be produced from the genetically altered protein; and b) producing in silico peptide fragments of the pre-defined length at a site of alteration in the protein comprising at least one mutated amino acid of the genetically altered protein.

8. The method of claim 1, wherein the length of the peptide fragments to be produced from the genetically



or equal to -5.5 using a ROBB760108 helix/turn scale.

17. The method of claim 1, wherein in step iii), the peptide variants so selected have: (a) at positions 5 and 6 an average value greater than 0.5 using RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn motif scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value greater than -0.34 using a FNSA.2 charge of side chain scale, (e) at positions 6 and 7 an average value greater than -5.5 using a ROBB760108 helix/turn scale, (f) at positions 1-9 an average value less than or equal to 45.56 using a NAKH920106 helix/turn motif scale, and (g) at positions 2 and 3 an average value greater than -0.055 using a QIAN880139 helix/turn motif scale.

18. The method of claim 1, wherein in step iii), the peptide variants so selected have: (a) at positions 5 and 6 an average value greater than 0.65 using RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn motif scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value greater than -0.34 using a FNSA.2 charge of side chain scale, (e) at positions 6 and 7 an average value greater than -5.5 using a ROBB760108 helix/turn scale, (f) at positions 1-9 an average value greater than 45.56 using a NAKH920106 helix/turn motif scale, and (g) at positions 2 and 3 an average value greater than -0.055 using a QIAN880139 helix/turn motif scale, (h) at positions 7 and 8 an average value less than or equal to -0.23 using a QIAN880138 helix/turn motif scale, and (i) at positions 1-9 an average value greater than 7.0 using a CHAM830103 steric scale.

19. The method of claim 1, wherein in step iii), the peptide variants so selected have: (a) at positions 5 and 6 an average value greater than 0.5 using RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn motif scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value greater than -0.34 using a FNSA.2 charge of side chain scale, (e) at positions 6 and 7 an average value greater than -5.5 using a ROBB760108 helix/turn scale, (f) at positions 1-9 an average value greater than 45.56 using a NAKH920106 helix/turn motif scale, (g) at positions 7 and 8 an average value greater than -0.23 using a QIAN880138 helix/turn motif scale, (h) at positions 1 and 2 an average value less than or equal to 0.625 using a MITS020101 solvent accessibility of an amino acid scale, (i) at position 2 a value less than or equal to 0.144401 using PNSA.1.AUTO charge of side chain scale, (j) at position 2 a value greater than -0.303435 using PNSA.1.AUTO charge of side chain scale, (k) at position 3 a value less than or equal to 6.8 using a KARS160118 amino frequency scale, and (l) at positions 8 and 9 an average value less than or equal to 18.04 using a YUTK870104 hydrophobic scale.

20. The method of claim 1, wherein in step iii), the peptide variants so selected have: (a) at positions 5 and 6 an average value greater than 0.5 using RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn motif scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value greater than -0.34 using a FNSA.2 charge of side chain scale, (e) at positions 6 and 7 an average value greater than -5.5 using a ROBB760108 helix/turn scale, (f) at positions 1-9 an average value greater than 45.56 using a NAKH920106 helix/turn motif scale, (g) at positions 7 and 8 an average value greater than -0.23 using a QIAN880138 helix/turn motif scale, (h) at positions 1 and 2 an average value less than or equal to 0.625 using a MITS020101 solvent accessibility of an amino acid scale, (i) at positions 2 a value less than or equal to 0.144401 using a PNSA.1.AUTO charge of side chain scale, (j) at position 3 a value greater than 6.8 using a KARS160118 amino frequency scale, and (k) at positions 5 and 6 an average value less than or equal to 17.92 using a YUTK870103 hydrophobic scale.

21. The method of claim 1, wherein in step iii), the peptide variants so selected have a helix/turn motif (a) at positions 5 and 6 an average value greater than 0.5 using RICJ880105 helix/turn motif scale, (b) at positions 1, 2, 8, and 9 an average value greater than -0.77 using a QIAN880107 helix/turn motif scale, (c) at positions 8 and 9 an average value greater than 17.75 using a YUTK870103 hydrophobic scale, (d) at position 3 a value greater than -0.34 using a FNSA.2 charge of side chain scale, (e) at positions 6 and 7 an average value greater than -5.5 using a ROBB760108 helix/turn scale, (f) at positions 1-9 an average value greater than 45.56 using a NAKH920106 helix/turn motif scale, (g) at positions 7 and 8 an average value greater than -0.23 using a QIAN880138 helix/turn motif scale, (h) at positions 1 and 2 an average value less than or equal to 0.625 using a MITS020101 solvent accessibility of an amino acid scale, and (i) at position 2 a value





scale applied to positions 6 and 7 of the peptide, nnn. PONP800103 hydrophobic scale applied to positions 6 and 7 of the peptide, ooo. KANM800104 hydrophobic scale applied to positions 1, 2, 8 and 9 of the peptide, ppp. ZASB820101 hydrophobic scale applied to positions 1, 2, 3, 4, 5, 6, 7, 8 and 9 of the peptide, qqq. WILM950103 hydrophobic scale applied to positions 1, 2, 3, 4, 5, 6, 7, 8 and 9 of the peptide, rrr. YUTK870103 hydrophobic scale applied to positions 5 and 6 of the peptide, sss. YUTK870103 hydrophobic scale applied to positions 8 and 9 of the peptide, and ttt. YUTK870104 hydrophobic scale applied to positions 8 and 9 of the peptide, or a combination thereof.

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### *Description*

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#### FIELD OF THE DISCLOSURE

The present disclosure is directed to methods of identifying immunogenic mutant peptides having therapeutic utility as cancer vaccines

#### BACKGROUND OF THE INVENTION

Genetic alterations are detected in all tumor cells. These alterations, occurring at the level of DNA, are transcribed and translated to generate altered proteins that in many instances drive cancer. These altered proteins can sometime contribute to immune recognition by T and B cells evoking activation of the immune response, which can lead to the elimination of tumor cells expressing the altered proteins [1-3].

Tumor cells, including malignant tumor cells or cancer cells, accumulate a large number of somatic mutations, from as low as ten, to as high as thousands depending on the cancer type. Only a subset of these mutations can evoke an immune response. Identifying such mutations can lead to the generation of therapeutic vaccines that can be given to patient as a polypeptide or as nucleic acids (both DNA and RNA) [4].

For a mutation to be recognized as foreign, the mutant amino acid should be present as part of a peptide that binds class I or class II major histocompatibility complex (MHC or alternatively known as human leukocyte antigen or HLA in human) molecules and be presented on the surface of antigen presenting cells (professional APCs). The MHC- or HLA-bound peptide interacts with the T-cell receptor (TCR) expressed on the surface of T cells. Productive binding with the TCR activates T-cells, which can kill tumor cells directly through its cytolytic activity (CD8<sup>+</sup> cytotoxic T-cells) or perform helper function (CD4<sup>+</sup> helper T-cells) to induce antibody production. In this context, the definition of an immunogenic peptide is restricted to peptides that can interact with CD8<sup>+</sup> or CD4<sup>+</sup> T cells. For the interaction to happen, the peptide must be presented on the surface of cells in complex with MHC or HLA class I or class II proteins. The MHC class I- or HLA class I-bound peptide interacts with CD8<sup>+</sup> T cells, and the MHC class II- or HLA class II-bound peptide interacts with CD4<sup>+</sup> T cells. Although MHC or HLA binding and surface presentation is required for T cell activation, but, the displayed peptide bound to MHC or HLA proteins on the surface of cell is necessary but not sufficient for T cell activation as TCR must also interact with the displayed peptide. Most peptides presented on the cell surface in complex with MHC or HLA fail to engage T cells and therefore are not immunogenic [5]. Immunogenicity require not only peptide-binding and display by MHC class I or class II proteins but also binding of the MHC class I or class II-displayed peptide by TCR of the CD8<sup>+</sup> T-cell or CD4<sup>+</sup> T-cell, respectively [6]. While much is known about the rules governing peptide binding by MHC or HLA molecules, little is known about the rules governing peptide binding by TCR, other than that the rules governing peptide binding by TCR are different from peptide binding by MHC or HLA proteins.

Class I HLA proteins are encoded by HLA-A, HLA-B and HLA-C genes. These proteins bind peptides of 8-11 amino acids in length, with the preferred length being 9 amino acids long. The peptide binding groove of class HLA is formed by two alpha helices supported by an anti-parallel beta sheet. The peptide-binding groove is deeper compared to class II HLA molecules and requires residues to be projected outside the binding groove to make interactions with the TCR [7].

Peptides bind to class HLA molecules in a multistep process. The steps are as follows: 1) generation of

protein fragments by immunoproteasomal or proteasomal processing as part of the natural turnover of proteins in cells [8]; 2) Entry of the protein fragment into the lumen of the endoplasmic reticulum by binding to peptide transporters (TAP) [9]; 3) Binding to the peptide-binding groove of the class I HLA molecules; 4) Transport through vesicles to the cell surface and 5) presentation on the surface of cells [10] [11].

In the case of endogenous proteins, such as altered proteins in tumor or cancer cells, these proteins being produced intracellularly by the cell do not require cellular uptake. As such, peptides derived by immunoproteasomal or proteasomal processing as part of the natural turnover of proteins in cells may be displayed by class I MHC or HLA molecules in all cell types in which the altered protein is expressed by the cell. In contrast, in the case of a peptide used in tumor or cancer vaccine, the peptide is exogenous to the cell and must be taken up by professional antigen-presenting cells in a process called cross-presentation in order to be displayed by class I MHC or HLA proteins [12-14]. The peptide used in tumor or cancer vaccine is longer than the peptide displayed by class I MHC or HLA proteins, as the peptide is taken up by the cell and undergo proteolysis to produce shorter peptide(s). Equal number of amino acids are added to the amino- and carboxy-termini, so as to extend the length of the final peptide displayed by class I MHC or HLA proteins. Typically, five to eighteen amino acids are added to each end of the 8-11 amino acid long peptide displayed on cell surface by class I MHC or HLA proteins, such that the peptide formulated in the tumor or cancer vaccine is approximately 18 to 47 amino acids in length. The upper limit of peptide length in tumor or cancer vaccine is less than or equal to 50 amino acids. The antigen-presenting cells capable of cross presentation are professional antigen-presenting cells and include dendritic cells (primarily), macrophages, and B lymphocytes.

The binding of MHC-peptide complex to the CD8<sup>sup.</sup>+ T cells, henceforth referred to as cytolytic or cytotoxic T cells (CTLs) activates a series of signaling pathways in CTLs resulting in their expansion to generate a population of effector CTLs. These CTLs will recognize tumor cells displaying the mutant peptide on their surface and kill them by apoptosis. Therefore, peptides derived from cancer mutations that are capable of mounting a CTL response can be used as cancer vaccines for treating cancer patients [15].

Two studies have demonstrated that immunogenic peptides can provide long term benefit to cancer patients when used as monotherapy [16, 17]. Therefore, accurate identification of immunogenic peptides from tumor-derived mutant protein can provide an avenue of treatment for cancer patients [18] [19]. However, the lack of efficient method for identifying bonafide immunogenic peptides have not only increased the cost of vaccination, but also increased the uncertainty of whether the vaccine will deliver the desired effect of inducing an anti-tumor response.

Next generation sequencing technology can catalogue all tumor mutations from a patient's tumor cells rapidly. However, identifying immunogenic peptides derived from such mutations is still a formidable challenge. The challenge comes from the fact that accurate methods of selecting immunogenic peptides from a pool of immunogenic and non-immunogenic peptides [20] [18].

Most screening platform uses HLA-binding prediction as a measure of immunogenicity [21]. The prediction can be further confirmed by actual detection of the peptide on the cell surface by mass spectrometry [5]. However, surface presentation of a peptide in complex with HLA is not an indication of immunogenicity. For a peptide to be immunogenic, the peptide presented on the surface of cells must engage T cell receptor. There is a need in the art for a high throughput methodology for prediction of immunogenic peptide for cancer therapy.

## SUMMARY OF THE DISCLOSURE

The practice matter of the invention disclosed in this application has employed, unless otherwise indicated, computational prediction algorithms organized in a step-wise workflow to identify tumor or cancer vaccines from tumor-derived proteins, which are expressed and mutated or altered only in cancer cells. The invention covers the identification of T-cell neo-epitopes from four classes of genetically altered proteins--i) proteins altered in amino acid sequence in which one or more amino acids are altered or mutated, which may be arranged in a sequence or distributed randomly across the length of the protein; ii) proteins produced from genes with internal insertion or deletion in the coding sequence; iii) proteins translated from fusion genes; and iv) proteins produced from splice variants.

Selection of immunogenic peptides comprises: a) selecting a set of cancer variants from mouse and human cancer cell lines and mouse and human cancer tissues where each variant in the genomic sequence correspond to both protein coding and protein non-coding sequences; b) variants of mouse cell lines and cancer tissues are identified by mouse whole exome and/or whole genome sequencing and variants from human cancer cell lines and human cancer tissues are identified by whole exome and/or whole genome sequencing; c) variants in mouse tissues and cell lines are identified by comparing with the reference sequence of mouse, and variants in human tissues and cell lines are identified by comparing with the reference sequence of human; d) variants are identified by comparing with the reference sequence, where the reference sequence is mouse reference sequence available in the public domain, or human reference sequence available in the public domain (e.g., current mouse reference sequence is (GRCm38/mm10) and current human reference sequence is (hg19)); e) variants from mouse tissues and cell lines include all genomic variants that alter the sequence of the RNA and the sequence of the protein translated from the RNA; f) variants from human tissues and cell lines include all genomic variants that alter the sequence of the proteins translated from the messenger RNA--protein variants; g) selecting the variants based on their expression in the mouse or human cell lines and tissues from the transcriptomic analysis; h) generating 8-11 amino acid peptides from the altered protein variants; and i) selecting a set of 8-11 amino acid immunogenic peptides from the previous step by predicting immunogenicity of the variant peptide comprising the altered amino acids encoded by the variant coding sequence; thereby selecting immunogenic peptides from altered or mutated proteins unique to cancer or tumor cells or tissues.

In some embodiments, according to any of the methods described above, the method further comprises selecting peptides that bind T cells by engaging with the T cell receptor (TCR) by obtaining peptides that carry features of TCR binding. Steps include one or more of: a) determining features associated with each of the amino acids in a 9-mer peptide; b) determining features that are unique or shared between amino acids that make up the composition of the 9-mer peptide; c) determining features that favor interactions between TCR and the HLA-bound peptide, comprising amino acid positions 3-8 of the 9-mer peptide; d) determining features that favor HLA binding comprising amino acid positions 1-2 and 9 of the 9-mer peptide; e) determining features that are different between the non-mutated and the mutated peptide; g) determining and/or applying features that select immunogenic peptides from a list of immunogenic and non-immunogenic peptides thereby identifying immunogenic peptides from altered proteins expressed in tumor or cancer cell lines and/or tissues.

According to any one of the methods described above immunogenic peptide is defined by a combination of one or more of the following parameters: i) peptide is derived from a gene which is mutated in the DNA from tumor or cancer cell but not in normal cell as determined by DNA sequencing; ii) the mutant gene is expressed in tumor or cancer and detected by transcriptome sequencing; iii) mutation changes one or more amino acids in the translated protein determined by in silico protein translation (conceptual translation of protein coding region or sequences) from the transcript encoding the mutant protein; iv) mutated or altered peptide derived from the mutant or altered protein binds TCR; v) affinity of mutated peptide to class I HLA or equivalent; vi) sensitivity of the peptide to processing by proteasomal and/or immunoproteasomal enzymes and vii) ability of the peptide to bind peptide transporter present on the endoplasmic reticulum. In some embodiments, predicting immunogenicity is further based on HLA-typing analysis.

The present application in another aspect also provides tumor-specific immunogenic peptides identified by any of the above methods or combination of methods from human tumor patients. In some embodiments, the composition comprises of two or more tumor specific immunogenic mutant peptides described herein. In some embodiments, the composition further comprises an adjuvant

The present application in another aspect also provides cancer-specific immunogenic peptides identified by any of the above methods or combination of methods from human cancer patients. In some embodiments, the composition comprises of two or more cancer specific immunogenic mutant peptides described herein. In some embodiments, the composition further comprises an adjuvant

The present application in yet another aspect provides a method of creating an immunogenic composition comprising at least one tumor or cancer specific mutant peptide or a larger precursor encoding the 8- to 11-mer mutant immunogenic peptide identified by any of the methods described herein. In one embodiment, the method of creating an immunogenic composition comprises at least one tumor specific mutant peptide or a larger precursor encoding the 9-mer immunogenic peptide identified by any of the methods described herein.

In some embodiments, the immunogenic composition contains two or more immunogenic tumor-specific mutant peptides. In some embodiments, the immunogenic composition contains two or more immunogenic cancer-specific mutant peptides.

The present application also provides an immunogenic composition comprising at least one nucleic acid encoding tumor or cancer specific immunogenic peptide, or one nucleic acid encoding a larger precursor containing the 9-mer mutant immunogenic peptide identified by any of the methods described herein. In some embodiments, the immunogenic composition comprising a nucleic acid encoding two or more (up to about 20) tumor-specific mutant immunogenic peptides. In some embodiments, the immunogenic composition comprising a nucleic acid encoding two or more (up to about 20) cancer-specific mutant immunogenic peptides. In other embodiments, the immunogenic composition can be composed of a mixture of immunogenic peptides, or a DNA encoding one or more immunogenic peptides, or a RNA encoding one or more immunogenic peptides.

## BRIEF DESCRIPTION OF THE FIGURES

FIG. 1. Steps to identify immunogenic peptides from cancer tissues.

FIG. 2. Steps for the creation of classification models for predicting TCR-binding peptides derived from normal and cancer tissues.

FIG. 3a-b. (a) Binding affinity distribution of immunogenic and non-immunogenic peptides, (b) Distribution of peptide with  $\geq 500$  nM and  $< 500$  nM.

FIG. 4. A schematic of the steps used for creating the classification models to separate TCR-binding peptides (immunogenic) from those that did not bind TCR (non-immunogenic).

FIG. 5a-b. (a) Sensitivity and specificity of the 500 training/test instances using J4.8 classification approach, (b) ROC curve from the ensemble classifier.

FIG. 6a-b. (a) Sensitivity and specificity of the 433 classifier instances using J4.8 classification approach, (b) The ROC curve for the 433 classifiers (colored in RED), 45 classifiers (colored in Blue).

FIG. 7a-c. Features to identify selected peptides. (a) Number of features that define occupancy of amino acids at each position of the 9-mer peptide. (b) Number of features that define hydrophobicity and helix/turn properties of amino acids. (c) Enrichment of amino acids with helix-turn and hydrophobicity properties at each position of the 9-mer peptides.

FIG. 8. Shows a schematic representation of the assay.

FIG. 9. The data presented here shows a validated neoantigen restricted to HLA-A\*02.01 as evidenced by elevated levels of CD8 T cell activation markers, INF- $\gamma$ . and CD69 in flow cytometric based assays. Naive human CD8 T cells specific for the HLA-A\*02.01-restricted epitopes showed a positive response to a colorectal cancer derived mutant peptide over a wild-type (control) peptide when stimulated with peptide-pulsed allogeneic DCs. Melan-A (26-35 L, positive control) is used as a positive control.

## DETAILED DESCRIPTION OF THE INVENTION

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as is commonly understood by one of ordinary skill in the art to which this invention belongs. All patents, applications, published applications and other publications referred to herein are incorporated by reference in their entirety.

As used in the description of the invention and the appended claims, the singular forms "a", "an" and "the" are used interchangeably and intended to include the plural forms as well and fall within each meaning, unless the context clearly indicates otherwise. Also, as used herein, "and/or" refers to and encompasses any and all possible combinations of one or more of the listed items, as well as the lack of combinations when interpreted in the alternative ("or").





esophageal cancer, eye cancer, intraocular, retinoblastoma, metastatic melanoma, gallbladder cancer, gastric cancer, gastrointestinal carcinoid tumor, gastrointestinal stromal tumors, glioblastoma, glioma, hairy cell leukemia, head and neck cancer, hepatocellular carcinoma, hepatoma, Hodgkin lymphoma, hypopharyngeal cancer, Langerhans cell histiocytosis, laryngeal cancer, lip and oral cavity cancer, liver cancer, lobular carcinoma in situ, lung cancer, non-small cell lung cancer, small cell lung cancer, lymphoma, AIDS-related lymphoma, Burkitt lymphoma, non-Hodgkin lymphoma, cutaneous T-cell lymphoma, melanoma, squamous neck cancer, mouth cancer, multiple myeloma, myelodysplastic syndromes, myelodysplastic/myeloproliferative neoplasms, nasal cavity and paranasal sinus cancer, nasopharyngeal cancer, neuroblastoma, oral cavity cancer, oropharyngeal cancer, osteosarcoma, ovarian cancer, pancreatic carcinoma, papillary carcinomas, parathyroid cancer, pharyngeal cancer, pheochromocytoma, pineal parenchymal tumors, pineoblastoma, pituitary tumor, pleuropulmonary blastoma, primary central nervous system lymphoma, prostate cancer, rectal cancer, renal cell cancer, salivary gland cancer, sarcoma, Ewing sarcoma, soft tissue sarcoma, squamous cell carcinoma, Sezary syndrome, skin cancer, Merkel cell carcinoma, testicular cancer, throat cancer, thymoma, thymic carcinoma, thyroid cancer, urethral cancer, endometrial cancer, uterine cancer, uterine sarcoma, vaginal cancer, vulvar cancer, Waldenstrom macroglobulinemia, and Wilms tumor. In one embodiment, the tumor is a glioma. In one embodiment, the tumor is a tumor other than a glioma.

For example, an inhibition of growth of a cancer cell means that the rate of growth of a cancer cell that has been treated with a peptide of the invention is 5-fold, 10-fold, 20-fold, 30-fold, 40-fold, 50-fold, 100-fold, or more, less than that of a cancer cell that has not been treated with a peptide of the invention. As used herein, "inhibition" as it refers to the rate of growth of a cancer cell that has been treated with a peptide of the invention also means that the rate is 90%, 80%, 70%, 60%, 50%, 40%, 30%, 25%, 20%, 15%, 10%, 5% or less, lower than the rate of growth of a cancer cell that has not been treated with a peptide of the invention.

An inhibition of growth of a cancer cell also means that the number or growth of cancer cells that have been treated with a peptide of the invention is 5-fold, 10-fold, 20-fold, 30-fold, 40-fold, 50-fold, 100-fold, or more, less than the number or growth of cancer cells that have not been treated with a peptide of the invention. As used herein, "inhibition" as it refers to the rate of growth of a cancer cell also means that the number or growth of cancer cells that have been treated with a peptide of the invention is 90%, 80%, 70%, 60%, 50%, 40%, 30%, 25%, 20%, 15%, 10%, 5% or less, lower than the growth or number of cancer cells that have not been treated with a peptide of the invention.

As used herein, "cancer" may be used interchangeably with "tumor," and vice versa, except when expressly or inherently prohibited. Similarly, "MHC" may be used interchangeably with "HLA," and vice versa, except when expressly or inherently prohibited.

The term "unmutated or wild-type peptide" refers to a peptide derived from normal or healthy tissue cells or tissue. Normal or healthy cells or tissue are free of disease, and in the context of the invention, free of tumor/cancer tissue or cells. Unlike cancer-specific mutant peptide, tumor peptide variant(s) or cancer peptide variant(s), which are mutant or altered peptide specific to cancer or tumor cells or tissues and not present in non-tumor/cancer cells or tissue, the "unmutated or wild-type peptide" may be present in cancer or tumor cells or tissue.

As used herein, the terms "comprising" or "comprises" is intended to mean that the compositions and methods include the recited elements, but not excluding others. "Consisting essentially of" when used to define compositions and methods, shall mean excluding other elements of any essential significance to the combination for the stated purpose. Thus, a composition consisting essentially of the elements as defined herein would not exclude other materials or steps that do not materially affect the basic and novel characteristic(s) of the present disclosure. "Consisting of" shall mean excluding more than trace elements of other ingredients and substantial method steps. Embodiments defined by each of these transition terms are within the scope of the present disclosure.

## Methods of the Invention

The invention describes a method for identifying immunogenic peptides from all genetically altered proteins derived from mammalian cancer samples using a high throughput approach. An accurate high throughput platform for the detection of immunogenic epitopes is critical for clinical translation. The immunogenic

peptides can be administered as personal cancer vaccines to individuals affected by the disease in the form of peptides, or as nucleotide-based precursors (e.g., DNA or RNA). The immunogenic peptides can have other applications in identifying specific TCR sequences that engage with the peptide, leading to the development of engineered T cells or CAR-T cells. Additionally, the immunogenic peptides can be used for developing TCR-mimetic reagents to target tumor cells. The methods described herein are useful in personalized cancer immunotherapy space for the treatment of individual cancer patients.

Thus, the present invention in one aspect provides a method of identifying cancer-specific mutant immunogenic peptide from the disease tissue of the individual by combining sequence-specific variant detection method with methods to determine immunogenicity of the peptides.

In another aspect, the present invention provides a method of identifying cancer-specific immunogenic peptides that bind T-cell receptor (TCR).

Also provided are enablement steps useful to practice the invention. Further included are a list of immunogenic peptides from cancer mutations detected by next generation sequencing, cancers presenting such peptides and nucleic acids encoding such peptides identified.

The invention provides methods of selecting cross species cancer vaccines from genetically altered proteins expressed by mouse and human cancer cells and/or tissues. In one embodiment, the method comprises (a) calculating the probability of HLA binding with optimal processing sites from a library of mutant cancer peptides; (b) calculating the probability of TCR binding to generate a T-cell response; and selecting the mutant cancer peptides having the highest probability so calculated from step (a) that can modulate the immune response of a mouse and a human, when challenged with the mutant cancer peptide thereby selecting cross species cancer vaccines; wherein the mouse and human subjects carry the same mutation and express the same HLA molecule that binds the mutant cancer peptide.

In accordance with the practice of the invention the tumor may be derived from any cancer. Examples of cancer cells or tissues include, but are not limited to, cancers of the Breast, Lung, Head & Neck, Skin, Ovary, Pancreatic, Liver, Brain, Prostate, Cervical Thyroid, Bone and Stomach.

The invention further provides methods of selecting mammalian tumor vaccine(s) from genetically altered protein(s) expressed by a mammalian tumor cell or a mammalian tumor tissue from a subject. In one embodiment of the invention, the method comprises the step of obtaining a sample from the subject. The sample may be directly processed as soon as it is obtained or the sample may be stored for a period of time before it is processed in accordance with the invention. The sample obtained from the subject may be cultured in vitro or used to produce cell line before processing in accordance with the invention. The method further comprises the step of identifying the genetically altered protein(s) expressed by the mammalian tumor cell or the mammalian tumor tissue in the sample through nucleic acid sequence(s) encoding the altered protein(s). Additionally, the method includes the step of producing peptide fragment(s) comprising at least one amino acid mutation from the genetically altered protein(s) so identified, so as to obtain peptide variant(s) associated with the mammalian tumor cell or the mammalian tumor tissue. In one embodiment, the peptide fragments are produced in silico using a sliding window method for a fixed or defined peptide length with one amino acid step producing a series of overlapping peptides of a pre-defined length with any mutant amino acid occupying different amino acid position in the series of peptides produced by the sliding window method.

Further, the method additionally comprises the step of selecting the peptide variant(s) which binds T-cell receptor (TCR). In one embodiment, this step comprises i) selecting the peptide variant(s) with a pre-defined length; ii) characterizing the peptide variant(s) in silico by selecting and matching features associated with an amino acid at each position of the peptide with selected pre-defined features for each position of peptides recognized by TCR associated with either CD8<sup>+</sup> T-cell or CD4<sup>+</sup> T-cell, so as to obtain predictive ability of the peptide variant(s) to interact with the TCR; iii) selecting the peptide variant(s) in step (ii) based on predicted ability of the peptide variant(s) to interact with the TCR, so as to be an immunogenic peptide that may or can serve as a mammalian tumor vaccine(s). Basis for mammalian tumor vaccine(s) using peptide variant(s) identified and selected by the methods of the invention require lengthening the selected peptide variant(s) such that following vaccination the lengthened selected peptide variant(s) is taken up by antigen-presenting cells, processed to the size of the selected peptide variant(s) (before lengthening) and displayed by







protein or the peptide fragment(s) of the pre-defined length is about 9, 10 or 11 amino acids long. In a specific example, the length of the peptide fragment(s) to be produced from the genetically altered protein or the peptide fragment(s) of the pre-defined length is 9 amino acids long.

In yet another embodiment, the length of the peptide fragment(s) further supports interaction with the TCR of CD8+ T-cell or CD4+ T-cell.

In still another embodiment, the interaction with the TCR of CD8+ T-cell or CD4+ T-cell results in a complex comprising the peptide, MHC class I protein and TCR of CD8+ T-cell, or alternatively, the peptide, MHC class II protein and TCR of CD4+ T-cell.

In an additional embodiment, interaction with the TCR of CD8+ T-cell or CD4+ T-cell results in a complex comprising the peptide, MHC class I protein and TCR of CD8+ T-cell, or alternatively, the peptide, MHC class II protein and TCR of CD4+ T-cell.

Also, in another embodiment, the mammalian tumor cell is a cell of a mammalian cell line derived from the tumor of a mammal. Merely by way of example, the mammal is selected from the group of human, mouse, rat, cat, dog, bovine, pig, sheep, goat, cow, horse, hamster, guinea pig, rabbit, mink, monkey, chimpanzee, and ape. In one embodiment, the mammal is a mouse or a human. In another embodiment, the tumor is a cancer. In yet a further embodiment, the mammalian tumor cell is a cell of a mouse cancer cell line. In a further still embodiment, the mammalian tumor cell is a cell of a human cancer cell line. Further, the mammalian tumor cell or mammalian tumor tissue may be present in or derived from a mouse or human subject.

Additionally, in accordance with the practice of the invention, the features associated with an amino acid at each position of the peptide may be physicochemical and/or biological properties of the amino acid. For example, each physicochemical and/or biological property of an amino acid may be assigned a numerical value within the context of other numerical values assigned to other amino acids.

Suitable examples of pre-defined features in accordance with the invention, include, but are not limited to, one or more of alpha-CH chemical shifts, hydrophobicity index (1), signal sequence helical potential, membrane-buried preference parameters, conformational parameter of inner helix, conformational parameter of beta-structure, conformational parameter of beta-turn, average flexibility indices, residue volume, information value for accessibility--average fraction 35%, information value for accessibility--average fraction 23%, retention coefficient in TFA, retention coefficient in HFBA, transfer free energy to surface, apparent partial specific volume, alpha-NH chemical shifts, alpha-CH chemical shifts, spin-spin coupling constants  $3J_{\text{H}\alpha\text{-NH}}$ , normalized frequency of alpha-helix, normalized frequency of extended structure, steric parameter, polarizability parameter, free energy of solution in water-kcal/mole, Chou-Fasman parameter of the coil conformation, a parameter defined from the residuals obtained from the best correlation of the Chou-Fasman parameter of beta-sheet, number of atoms in the side chain labelled 1+1, number of atoms in the side chain labelled 2+1, number of atoms in the side chain labelled 3+1, number of bonds in the longest chain, a parameter of charge transfer capability, a parameter of charge transfer donor capability, average volume of buried residue, residue accessible surface area in tripeptide, residue accessible surface area in folded protein, proportion of residues 95% buried, proportion of residues 100% buried, normalized frequency of beta-turn-1, normalized frequency of alpha-helix, normalized frequency of beta-sheet, normalized frequency of beta-turn-2, normalized frequency of N-terminal helix, normalized frequency of C-terminal helix, normalized frequency of N-terminal non helical region, normalized frequency of C-terminal non helical region, normalized frequency of N-terminal beta-sheet, normalized frequency of C-terminal beta-sheet, normalized frequency of N-terminal non beta region, normalized frequency of C-terminal non beta region, frequency of the 1st residue in turn, frequency of the 2nd residue in turn, frequency of the 3rd residue in turn, frequency of the 4th residue in turn, normalized frequency of the 2nd and 3rd residues in turn, normalized hydrophobicity scales for alpha-proteins, normalized hydrophobicity scales for beta-proteins, normalized hydrophobicity scales for alpha+beta-proteins, normalized hydrophobicity scales for alpha/beta-proteins, normalized average hydrophobicity scales, partial specific volume, normalized frequency of middle helix, normalized frequency of beta-sheet, normalized frequency of turn, size, amino acid composition, relative mutability, membrane preference for cytochrome b: MPH89, average membrane preference: AMP07, consensus normalized hydrophobicity scale, solvation free energy, atom-based hydrophobic moment, direction of hydrophobic moment, molecular weight, melting point, optical rotation, pK-N, pK-C,

hydrophobic parameter  $\pi$ , graph shape index, smoothed epsilon steric parameter, normalized van der Waals volume, STERIMOL length of the side chain, STERIMOL minimum width of the side chain, STERIMOL maximum width of the side chain, N.M.R. chemical shift of alpha-carbon, localized electrical effect, number of hydrogen bond donors, number of full nonbonding orbitals, positive charge, negative charge,  $pK_a(\text{RCOOH})$ , helix-coil equilibrium constant, helix initiation parameter at position  $i-1$ , helix initiation parameter (at position  $i$ ,  $i+1$ , and  $i+2$ ), helix termination parameter (at position  $j-2$ ,  $j-1$ , and  $j$ ), helix termination parameter at position  $j+1$ , partition coefficient, alpha-helix indices, alpha-helix indices for alpha-proteins, alpha-helix indices for beta-proteins, alpha-helix indices for alpha/beta-proteins, beta-strand indices, beta-strand indices for beta-proteins, beta-strand indices for alpha/beta-proteins, aperiodic indices, aperiodic indices for alpha-proteins, aperiodic indices for beta-proteins, aperiodic indices for alpha/beta-proteins, hydrophobicity factor, residue volume, composition, polarity, volume, partition energy, hydration number, hydrophilicity value, heat capacity, absolute entropy, entropy of formation, normalized relative frequency of alpha-helix, normalized relative frequency of extended structure, normalized relative frequency of bend, normalized relative frequency of bend R, normalized relative frequency of bend S, normalized relative frequency of helix end, normalized relative frequency of double bend, normalized relative frequency of coil, average accessible surface area, percentage of buried residues, percentage of exposed residues, ratio of buried and accessible molar fractions, transfer free energy, hydrophobicity (1),  $pK(-\text{COOH})$ , relative frequency of occurrence, relative mutability, amino acid distribution, sequence frequency, average relative probability of helix, average relative probability of beta-sheet, average relative probability of inner helix, average relative probability of inner beta-sheet, flexibility parameter for no rigid neighbors, flexibility parameter for one rigid neighbor, flexibility parameter for two rigid neighbors, Kerr-constant increments, net charge, side chain interaction parameter (1), side chain interaction parameter (2), fraction of site occupied by water, side chain volume, hydropathy index, transfer free energy, CHP/water, hydrophobic parameter, distance between C-alpha and centroid of side chain, side chain angle  $\theta(\text{AAR})$ , side chain torsion angle  $\phi(\text{AAAR})$ , radius of gyration of side chain, van der Waals parameter  $RO$ , van der Waals parameter  $\epsilon$ , normalized frequency of alpha-helix with weights, Normalized frequency of beta-sheet with weights, normalized frequency of reverse turn with weights, normalized frequency of alpha-helix (unweighted), normalized frequency of beta-sheet (unweighted), normalized frequency of reverse turn (unweighted), frequency of occurrence in beta-bends, conformational preference for all beta-strands, conformational preference for parallel beta-strands, conformational preference for antiparallel beta-strands, average surrounding hydrophobicity, normalized frequency of alpha-helix, normalized frequency of extended structure, normalized frequency of zeta R, normalized frequency of left-handed alpha-helix, normalized frequency of zeta L, normalized frequency of alpha region, refractivity, retention coefficient in HPLC ( $\text{pH}7.4$ ), retention coefficient in HPLC ( $\text{pH}2.1$ ), retention coefficient in  $\text{NaClO}_4$ , retention coefficient in  $\text{NaH}_2\text{PO}_4$ , average reduced distance for C-alpha, average reduced distance for side chain, average side chain orientation angle, effective partition energy, normalized frequency of alpha-helix, normalized frequency of beta-structure, normalized frequency of coil, AA composition of total proteins, SD of AA composition of total proteins, AA composition of mt-proteins, normalized composition of mt-proteins, AA composition of mt-proteins from animal, normalized composition from animal, AA composition of mt-proteins from fungi and plant, normalized composition from fungi and plant, AA composition of membrane proteins, normalized composition of membrane proteins, transmembrane regions of non-mt-proteins, transmembrane regions of mt-proteins, ratio of average and computed composition, AA composition of CYT of single-spanning proteins, AA composition of CYT2 of single-spanning proteins, AA composition of EXT of single-spanning proteins, AA composition of EXT2 of single-spanning proteins, AA composition of MEM of single-spanning proteins, AA composition of CYT of multi-spanning proteins, AA composition of EXT of multi-spanning proteins, AA composition of MEM of multi-spanning proteins, 8 A contact number, 14 A contact number, transfer energy, organic solvent/water, average non-bonded energy per atom, short and medium range non-bonded energy per atom, long range non-bonded energy per atom, average non-bonded energy per residue, short and medium range non-bonded energy per residue, optimized beta-structure-coil equilibrium constant, optimized propensity to form reverse turn, optimized transfer energy parameter, optimized average non-bonded energy per atom, optimized side chain interaction parameter, normalized frequency of alpha-helix from LG, normalized frequency of alpha-helix from CF, normalized frequency of beta-sheet from LG, normalized frequency of beta-sheet from CF, normalized frequency of turn from LG, normalized frequency of turn from CF, normalized frequency of alpha-helix in all-alpha class, normalized frequency of alpha-helix in alpha+beta class, normalized frequency of alpha-helix in alpha/beta class, normalized frequency of beta-sheet in all-beta class, normalized frequency of beta-sheet in alpha+beta class, normalized frequency of beta-sheet in alpha/beta class, normalized frequency of turn in all-alpha class, normalized frequency of turn in all-beta class, normalized frequency of turn in alpha+beta class, normalized frequency of turn in alpha/beta



of unfolding (pH7.0), activation Gibbs energy of unfolding (pH9.0), dependence of partition coefficient on ionic strength, hydrophobicity (3), bulkiness, polarity, isoelectric point, RF rank, normalized positional residue frequency at helix termini N4', normalized positional residue frequency at helix termini N''', normalized positional residue frequency at helix termini N'', normalized positional residue frequency at helix termini N', normalized positional residue frequency at helix termini Nc, normalized positional residue frequency at helix termini N1, normalized positional residue frequency at helix termini N2, normalized positional residue frequency at helix termini N3, normalized positional residue frequency at helix termini N4, normalized positional residue frequency at helix termini N5, normalized positional residue frequency at helix termini C5, normalized positional residue frequency at helix termini C4, normalized positional residue frequency at helix termini C3, normalized positional residue frequency at helix termini C2, normalized positional residue frequency at helix termini C1, normalized positional residue frequency at helix termini Cc, normalized positional residue frequency at helix termini C', normalized positional residue frequency at helix termini C'', normalized positional residue frequency at helix termini C''', normalized positional residue frequency at helix termini C4', Delta G values for the peptides extrapolated to 0 M urea, helix formation parameters (delta G), normalized flexibility parameters (B-values)--average, normalized flexibility parameters (B-values) for each residue surrounded by none rigid neighbors, normalized flexibility parameters (B-values) for each residue surrounded by one rigid neighbors, normalized flexibility parameters, Free energy in alpha-helical conformation, free energy in alpha-helical region, Free energy in beta-strand conformation, free energy in beta-strand region, free energy in beta-strand region, free energies of transfer of AcWL-X-LL peptides from bilayer interface to water, thermodynamic beta sheet

propensity, turn propensity scale for transmembrane helices, alpha helix propensity of position 44 in T4 lysozyme, p-Values of mesophilic proteins based on the distributions of B values, p-Values of thermophilic proteins based on the distributions of B values, distribution of amino acid residues in the 18 non-redundant families of thermophilic proteins, distribution of amino acid residues in the 18 non-redundant families of mesophilic proteins, distribution of amino acid residues in the alpha-helices in thermophilic proteins, distribution of amino acid residues in the alpha-helices in mesophilic proteins, side-chain contribution to protein stability (kJ/mol), propensity of amino acids within pi-helices, hydrophathy scale based on self-information values in the two-state model (5% accessibility), hydrophathy scale based on self-information values in the two-state model (9% accessibility), hydrophathy scale based on self-information values in the two-state model (16% accessibility), hydrophathy scale based on self-information values in the two-state model (20% accessibility), hydrophathy scale based on self-information values in the two-state model (25% accessibility), hydrophathy scale based on self-information values in the two-state model (36% accessibility), hydrophathy scale based on self-information values in the two-state model (50% accessibility), averaged turn propensities in a transmembrane helix, alpha-helix propensity derived from designed sequences, beta-sheet propensity derived from designed sequences, composition of amino acids in extracellular proteins (percent), composition of amino acids in anchored proteins (percent), composition of amino acids in membrane proteins (percent), composition of amino acids in intracellular proteins (percent), composition of amino acids in nuclear proteins (percent), surface composition of amino acids in intracellular proteins of thermophiles (percent), surface composition of amino acids in intracellular proteins of mesophiles (percent), surface composition of amino acids in extracellular proteins of mesophiles (percent), surface composition of amino acids in nuclear proteins (percent), interior composition of amino acids in intracellular proteins of thermophiles (percent), interior composition of amino acids in intracellular proteins of mesophiles (percent), interior composition of amino acids in extracellular proteins of mesophiles (percent), interior composition of amino acids in nuclear proteins (percent), entire chain composition of amino acids in intracellular proteins of thermophiles (percent), entire chain composition of amino acids in intracellular proteins of mesophiles (percent), entire chain composition of amino acids in extracellular proteins of mesophiles (percent), entire chain composition of amino acids in nuclear proteins (percent), screening coefficients gamma (local), screening coefficients gamma (non-local), slopes tripeptide--FDPB VFF neutral, slopes tripeptides--LD VFF neutral, slopes tripeptide--FDPB VFF noside, slopes tripeptide FDPB VFF all, slopes tripeptide FDPB PARSE neutral, slopes dekaepptide--FDPB VFF neutral, slopes proteins--FDPB VFF neutral, side-chain conformation by gaussian evolutionary method, amphiphilicity index, volumes including the crystallographic waters using the ProtOr, volumes not including the crystallographic waters using the ProtOr, electron-ion interaction potential values, hydrophobicity scales, hydrophobicity coefficient in RP-HPLC-C18 with 0.1% TFA/MeCN/H2O, hydrophobicity coefficient in RP-HPLC-C8 with 0.1% TFA/MeCN/H2O, hydrophobicity coefficient in RP-HPLC-C4 with 0.1% TFA/MeCN/H2O, hydrophobicity coefficient in RP-HPLC-C18 with 0.1% TFA/2-PrOH/MeCN/H2O, hydrophilicity scale, retention coefficient at pH 2, modified Kyte-Doolittle hydrophobicity scale, interactivity scale obtained from the contact matrix, interactivity scale obtained by



mammalian tumor tissue so selected. The peptide may be a peptide variant. Moreover, rank ordering peptides may be based on a combination of the following parameters: a) expression of variant gene from which variant peptide is derived; b) predicted ability to bind TCR of CD8+ T-cell; c) binding affinity of the peptide to MHC class-I protein(s); d) peptide processing by proteases; and/or e) peptide transporter binding. Further, each parameter may be subdivided to reflect quality of the parameter through numerical value(s) or range(s) of values, and further, the numerical value(s) or range(s) of values from the parameters assessed or combined so as to produce output(s) permissive of sorting by ascending or descending order, thereby predicting a rank ordered list of the immunogenic peptides derived from mammalian tumor cell or mammalian tumor tissue so selected.

In another embodiment, the methods of the invention may further comprise predicting a rank ordered list of immunogenic peptides derived from mammalian tumor cell or mammalian tumor tissue, wherein the peptide is a peptide variant and wherein rank ordering peptides is based on a combination of the following parameters: a) expression of variant gene from which variant peptide is derived; b) predicted ability to bind TCR of CD4+ T-cell; c) binding affinity of the peptide to MHC class-II protein(s); d) peptide processing by lysosome and/or endosome; and/or e) fusion of the endosomal and/or lysosomal vesicles with Golgi-derived vesicles to permit loading of the immunogenic peptide onto MHC class II proteins.

In one embodiment of the invention, the immunogenic peptide so selected may be further selected by its ability to bind MHC class-I or class-II protein(s) or for its ability to bind a specific MHC class-I protein derived from a particular allele of MHC class I gene or specific MHC class-II proteins derived from two particular MHC class II genes. For example, the MHC class-I or class-II protein(s) may be encoded by the human leukocyte antigen gene complex (HLA). As a further example, the particular allele of MHC class I gene may be encoded by HLA-A locus, HLA-B locus, HLA-C locus, HLA-E locus, HLA-F locus or HLA-G locus. Further examples of the particular allele of MHC class I gene may be selected from the set as shown in Table 2.

Additionally, in one embodiment, the specific MHC class-II proteins may be derived from two particular MHC class II genes to form a heterodimer of an alpha chain and a beta chain. For example, the heterodimer may be any or HLA-DM, HLA-DO, HLA-DP, HLA-DQ and HLA-DR. IN another example, the alpha chain of HLA-DM heterodimer may be encoded by HLA-DMA locus, alpha chain of HLA-DO heterodimer is encoded by HLA-DOA locus, alpha chain of HLA-DP heterodimer is encoded by HLA-DPA1 locus, alpha chain of HLA-DQ heterodimer is encoded by HLA-DQA1 locus or HLA-DQA2 locus, and alpha chain of HLA-DR is encoded by HLA-DR locus. In a further example, the beta chain of HLA-DM heterodimer may be encoded by any of HLA-DMB locus, beta chain of HLA-DO heterodimer is encoded by HLA-DOB locus, beta chain of HLA-DP heterodimer is encoded by HLA-DPB1 locus, beta chain of HLA-DQ heterodimer is encoded by HLA-DQB1 locus or HLA-DQB2 locus, and beta chain of HLA-DR is encoded by HLA-DRB1 locus, HLA-DRB3 locus, HLA-DRB4 or HLA-DRB5 locus. Further examples of the particular allele of MHC class II gene may be selected from the set as shown in Table 3.

In accordance with the invention the allele may be described by a classification system comprising HLA prefix, separated by hyphen, followed by HLA gene, field separator, serotype, protein coded by allele in order of discovery, one or more numbers designated by gene sequencing and expression, or a combination thereof. Currently, there are more than 7,670 MHC class I alleles and more than 2,260 MHC class II alleles. In addition, each locus may comprise multiple genes or alleles of MHC class-I or class-II protein(s).

In accordance with the invention, the methods of the invention may further comprise MHC-typing of the tumor cell or tumor tissue in selection of immunogenic peptide(s), so as to select immunogenic peptide(s) which may be displayed by the tumor cell or tumor tissue, by cells of individual or subject from which tumor cell or tumor tissue is derived, or by immune cells of individual or subject from which tumor cell or tumor tissue is derived.

In accordance with the invention, the methods of the invention may further comprise HLA-typing of the tumor cell or tumor tissue in selection of immunogenic peptide(s), so as to select immunogenic peptide(s) which may be displayed by the tumor cell or tumor tissue, by cells of individual or subject from which tumor cell or tumor tissue is derived, or by immune cells of individual or subject from which tumor cell or tumor tissue is derived.

In one embodiment of the invention, the prediction of immunogenic peptide(s) may further comprise MHC-typing analysis comprising the steps of: a) determining serotype or expressed isotype or supertype of MHC class-I or class-II protein(s) expressed by MHC class-I or class-II genes of the mammalian tumor cell or tumor tissue, or alternatively of the cell or immune cell of an individual or subject to be administered with mammalian tumor vaccine(s) comprising the predicted immunogenic peptide(s); b) calculating probability of MHC class-I or class-II protein(s) of (a) binding mammalian tumor peptide variant(s) with optimal processing sites from a library of tumor peptide variants; c) calculating probability of TCR binding to generate a T-cell response; d) selecting tumor peptide variant(s) having highest probability from steps (b) that can modulate the immune response of a mammal when challenged with the tumor peptide variant(s), thereby further selecting mammalian tumor vaccine(s) dependent on MHC class-I or class-II expression of the mammalian tumor cell or tumor tissue, or alternatively of the cell or immune cell of an individual or subject to be administered with mammalian tumor vaccine(s) comprising the predicted immunogenic peptide(s).

In another embodiment, the prediction of immunogenic peptide(s) may further comprise the steps of HLA-typing analysis comprising: a) determining serotype or expressed isotype or supertype of HLA protein(s) expressed by HLA genes of the mammalian tumor cell or tumor tissue, or alternatively of the cell or immune cell of an individual or subject to be administered with mammalian tumor vaccine(s) comprising the predicted immunogenic peptide(s); b) calculating probability of HLA protein(s) of (a) binding mammalian tumor peptide variant(s) with optimal processing sites from a library of tumor peptide variants; c) calculating probability of TCR binding to generate a T-cell response; d) selecting tumor peptide variant(s) having highest probability from steps (b) that can modulate the immune response of a mammal when challenged with the tumor peptide variant(s), thereby further selecting mammalian tumor vaccine(s) dependent on HLA expression of the mammalian tumor cell or tumor tissue, or alternatively of the cell or immune cell of an individual or subject to be administered with mammalian tumor vaccine(s) comprising the predicted immunogenic peptide(s).

In accordance with the invention, the mammalian tumor vaccine(s) may comprise the selected immunogenic peptide so identified by computation method.

Further, in accordance with the invention, selected immunogenic peptide in the mammalian tumor vaccine(s) may have amino-terminal and carboxyl-terminal extensions. For example, the amino-terminal and carboxyl-terminal extensions may be amino acids. The amino acids in the amino-terminal and carboxyl-terminal extensions may permit processing of the selected immunogenic peptide of claim 1 or 3 so as to be displayed by the MHC class I protein(s) and/or the MHC class II protein(s). For example, the MHC class I protein(s) and/or the MHC class II proteins(s) may be associated with a human. Further, the MHC class I protein(s) and/or the MHC class II protein(s) associated with a human may be an HLA protein(s).

Additionally, the invention provides methods of preparing a subject-specific immunogenic peptide composition comprising selecting cancer vaccines from genetically altered proteins expressed by mammalian cancer cells and tissues by any of the methods of the invention. Merely by way of example, said subject-specific peptides, may comprise: (a) a peptide that has a non-synonymous mutation leading to different amino acids in comparison with a protein of the non-tumor sample; (b) a peptide having a read-through mutation in which a stop codon is modified or deleted, leading to translation of a longer protein in comparison with a protein of the non-tumor sample with a novel tumor-specific sequence at the C-terminus; (c) a peptide that has a splice site mutation that leads to the inclusion of an intron or part of an intron, or alternatively exclusion of an exon or part of an exon, in the mature mRNA and thus has a unique tumor-specific protein sequence; (d) a peptide representing a chromosomal rearrangement that has given rise to a chimeric protein with tumor-specific sequences at the junction of two proteins of the non-tumor sample and thus represents a gene fusion; or (e) a peptide representing in comparison with a protein of the non-tumor sample a frameshift mutation or deletion that leads to a new open reading frame and a novel tumor-specific protein sequence. The subject-specific immunogenic composition may comprise a subject-specific peptide that binds to the HLA protein of the subject with an IC<sub>50</sub> less than about 500 nM.

The invention additionally provides methods of treating a subject having cancer. In one embodiment, the method comprises administering in the subject an immunogenic peptide, composition of the invention or cancer vaccines so selected by any of the methods of the invention in a sufficient amount so as to treat the cancer.

In another embodiment, the method comprises a) obtaining a sample from the subject; b) identifying the genetically altered protein(s) expressed by the mammalian tumor cell or the mammalian tumor tissue in the sample through nucleic acid sequence(s) encoding the altered protein(s); b) producing peptide fragment(s) comprising at least one amino acid mutation from the genetically altered protein(s) so identified in step (a), so as to obtain peptide variant(s) associated with the mammalian tumor cell or the mammalian tumor tissue. Then the method further comprises selecting the peptide variant(s) from step b, which binds a T-cell receptor (TCR). This step comprises: i) selecting the peptide variant(s) with a pre-defined length; ii) characterizing the peptide variant(s) (e.g. in silico) by selecting and matching features associated with an amino acid at each position of the peptide with selected pre-defined features for each position of peptides recognized by TCR associated with either CD8+ T-cell or CD4+ T-cell, so as to obtain predictive ability of the peptide variant(s) to interact with the TCR; iii) selecting the peptide variant(s) above based on predicted ability of the peptide variant(s) to interact with the TCR, so as to be an immunogenic peptide that may or can serve as a mammalian tumor vaccine(s) after lengthening the selected immunogenic peptide variant(s) such that following vaccination the lengthened selected peptide variant(s) is taken up by antigen-presenting cells, processed to the size of the selected peptide variant(s) and displayed by antigen-presenting cells. The method further comprises forming a vaccine comprising the at least one immunogenic peptide so selected and administering the vaccine in an effective amount to the subject so as to treat the cancer in the subject.

For example, the cancer may be a stomach cancer, a colon cancer, a breast cancer, an ovarian cancer, a prostate cancer, a lung cancer, a kidney cancer, a gastric cancer, a testicular cancer, a head and neck cancer, a pancreatic cancer, a brain cancer, a melanoma, a lymphoma or a leukemia.

#### Immunogenic Peptides from Mutated or Altered Proteins in Mammalian Cancers

The invention further provides an immunogenic peptide composition prepared by this method of the invention. In one embodiment, the immunogenic peptide composition may further comprise at least one adjuvant.

The invention further provides a mammalian tumor vaccine selected by any of the methods of the invention.

The methods described herein in various embodiments comprise identifying immunogenic peptides of nine amino acids (9-mer) derived from mutations present in mammalian cancer tissues and cancer cell lines. In the context of this disclosure, immunogenic peptides are selected on the basis of: i) TCR binding; ii) HLA binding; iii) expression; iv) proteolytic processing; and v) peptide transporter binding. The method described in various embodiments was applied to 2.3 million unique cancer mutations captured from MedGenome's proprietary cancer mutation database OncoMD.TM. and a list of peptides restricted to class I HLA molecules consisting of HLA-A01:01, HLA-A02:0, HLA-A11:01, HLA-A24:02, HLA-B35:03, HLA-B40:06, HLA-B44:03, HLA-B51:01, HLA-B57:01, HLA-C06:02, HLA-C07:02, HLA-C12:03, HLA-C15:02 are identified (Table 1). In some embodiments, one or more of the 9-mer immunogenic peptide identified by the methods of the invention can be used following amino acid extension (addition) on amino-terminus and carboxyl-terminus, as a cancer vaccine and administered to cancer patients. In an embodiment, equal number of amino acids are added at each end of the 9-mer peptide identified by the methods of the invention, so as to permit cross presentation of the desired 9-mer immunogenic peptide. In some embodiments, the composition of a cancer vaccine may comprise of two or more immunogenic peptides. In some embodiments, cancer vaccines comprising of one, two or more immunogenic peptides may activate a cytotoxic T cell (CTL) response and a CD4 T cell response against one or two or more immunogenic peptides.

In some embodiments, the cancer vaccine composition may comprise of a 9-mer immunogenic peptide that may be part of a precursor protein, or part of longer peptides about >9 amino acids up to about 50 amino acids. In some embodiments, the cancer vaccine composition may comprise of two or more immunogenic peptides that may be part of one, two or more precursor proteins or part of one, two or more longer peptides about >9 amino acids up to about 50 amino acids. In some embodiments, the composition of the cancer vaccine may contain an adjuvant to help boost the immune response. In some embodiments, the composition of the cancer vaccine containing an adjuvant to help boost the immune response may be pharmaceutically acceptable.

In some embodiments, the cancer vaccine, or a precursor protein containing the cancer vaccine, or a longer peptide about >9 amino acids up to about 50 amino acids containing the cancer vaccine may be encoded by a

nucleic acid sequence. In some embodiments, the nucleic acid sequence may be a DNA. In other embodiments, the nucleic acid sequence may be RNA. In some embodiments, the nucleic acid sequence may contain an adjuvant. In some embodiments, the nucleic acid sequence with the adjuvant may be used for treating the cancer patients.

In some embodiments, the nucleic acid sequence may be injected into mammalian cells to express the cancer vaccine in the form of a peptide, or as part of a protein precursor or as part of a longer peptide >9 amino acid up to about 50 amino acids to generate stable cells. In some embodiments, the stable cells may be primary cells, or cell lines derived from primary cells. In some embodiments, the primary cell may be derived from normal tissues or from cancer tissues.

In some embodiments, the stable cells may be used for screening antibodies by phage display technology. In some embodiments, the stable cells may be used in T cell activation screening assays.

### Combination Therapy

In another embodiment, the peptides of the invention (e.g., single or multiple peptides of the invention) so obtained by the methods of selection of the invention may be administered in combination, or sequentially, with another therapeutic agent. Such other therapeutic agents include those known for treatment, prevention, or amelioration of one or more symptoms of cancer diseases and disorders. Such therapeutic agents include, but are not limited to, ricin, ricin A-chain, doxorubicin, daunorubicin, taxol, ethidium bromide, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicine, dihydroxy anthracin dione, actinomycin D, diphtheria toxin, Pseudomonas exotoxin (PE) A, PE40, abrin, arbrin A chain, modeccin A chain, alpha-sarcin, gelonin, mitogellin, retstrictocin, phenomycin, enomycin, curicin, crotin, calicheamicin, sapaonaria officinalis inhibitor, maytansinoids, and glucocorticoid and other chemotherapeutic agents, as well as radioisotopes such as .sup.212Bi, .sup.131I, .sup.131In, .sup.90Y, and .sup.186Re.

The peptides of the invention formulated into tumor or cancer vaccine(s) may also be used in combination, or sequentially, with one or more immune checkpoint inhibitors. Immune checkpoint inhibitors include inhibitors for PD-1, PD-L1, PD-L2, 4-1BB, 4-1BBL, HVEM, BTLA, CD160, CD226, LAG3, CTLA-4, B7-1, B7-2, CD40, CD40L, Galectin-9, TIM-3, GITR, GITRL, SIRP alpha, B7-H3, B7-H4, VISTA, OX40, OX-40L, CEACAM1, CD47, ICOS, ICOSL, TIGIT, IDO, CD28, LIGHT, TIGIT, CD155, CD70 and adenosine A2a receptor. Immune checkpoint inhibitor may be an antibody or an antibody fragment. The antibody or antibody fragment may be derived from a monoclonal antibody. In one embodiment, the monoclonal antibody or its fragment is human or humanized. Immune checkpoint inhibitor for PD-1 may be selected from any of MEDI0680 (also known as AMP-614; MedImmune/AstraZeneca), nivolumab (also known as Opdivo, BMS-936558, MDX-1106 and ONO-4538; Bristol-Myers Squibb and Ono Pharmaceuticals), pembrolizumab (also known as Keytruda, MK-3475 and lambrolizumab; Merck) and pidilizumab (also known as CT-011; CureTech). Immune checkpoint inhibitor for PD-L1 may be selected from any of BMS-936559 (also known as CT-011; Bristol-Myers Squibb), MEDI4736 (MedImmune/AstraZeneca), MPDL3280A (also known as RG7446; Genetech/Roche) and MSB0010718C (EMD Serono).

### Kits

According to another aspect of the invention, kits are provided. Kits according to the invention include package(s) comprising antibodies or compositions of the invention.

The phrase "package" means any vessel containing peptides or compositions presented herein. In preferred embodiments, the package can be a box or wrapping. Packaging materials for use in packaging pharmaceutical products are well known to those of skill in the art. Examples of pharmaceutical packaging materials include, but are not limited to, blister packs, bottles, tubes, inhalers, pumps, bags, vials, containers, syringes, bottles, and any packaging material suitable for a selected formulation and intended mode of administration and treatment.

The kit can also contain items that are not contained within the package but are attached to the outside of the package, for example, pipettes.

Kits may optionally contain instructions for administering peptides or compositions of the present invention





peptide. When both sites are optimally located a maximum score of 20 is given. The score decreases as the processing sites are shifted away from the optimal location. A score  $>10$  is used to select peptides for the next step. Peptides that are scored higher than 10 either by the proteasomal or by the immunoproteasomal cleavage are selected.

Step 14 calculates the transporter (TAP) binding affinity of the peptides. In order for the peptide to bind HLA molecule, the peptide needs to be transported from cytosol to endoplasmic reticulum. In this step, we perform the analysis to identify whether the peptide is delivered to HLA molecule by TAP. Any peptide exhibiting a TAP-binding score of  $<0.5$  are selected for the final step of prioritization.

### Predicting Immunogenic Peptides by their Ability to Bind TCRs

The prediction of TCR-binding peptide prediction involves four different steps: 1. Data set creation; 2. Feature creation; 3. Classification model; 4. Study of features. The steps are shown in FIG. 2. A brief description of each step: 1. Dataset creation: In this step, we have first collected peptide and its immunogenicity status from IEDB database. After this we then performed processing of the peptides to have a clean dataset for the model building exercise. Further, we have generated several training and test instances for model building and performance evaluation. 2. Feature creation: In this step, various amino acid features, HLA binding and peptide processing related feature is generated for the peptides. 3. Classification model: In this step, classification model is generated using feature matrix. This step involves: feature selection, identification of classification method, scoring of the peptides. 4. Study of features: The important features are studied in detail and its correlation with peptide structure/interactions in crystal structure is also studied in this step.

### Data Preparation

The sequence, assay, HLA type, publication id (PMID), and immunogenicity information of the peptide was downloaded from IEDB database (Release 24 Nov. 2016). The database contains immunogenicity status for 2,521 unique 9-mer peptides for human. The peptide is first categorized into self and foreign peptide. The peptides generated by human body are known as self, while those that do not originate in human body are called non-self or foreign peptides. Of the total peptides, .about.85% of them belong to foreign peptide category. The peptides are also classified based on assay that was performed to check its immunogenicity. Although there are several assay types, we have broadly grouped them into biological and non-biological type. Majority of the peptides (.about.90%) are assayed by biological type. Before using these peptides, we apply the following filters to focus on unambiguous assay prediction and for which the information as per our requirement is complete. Biological assay filter: The peptides predicted as immunogenic/non-immunogenic using one of the biological assay is taken further for the analysis. Prediction by assays: There are many peptides which are predicted as both immunogenic and non-immunogenic using one or more different assays. These peptides were removed from our analysis. 4-digit HLA information: The peptides for which 4-digit information is available for the HLA type is considered for further analysis. Of the total peptides, for 1075 peptides 4-digit HLA information was available

Overall, we obtain 1,075 peptides for which unambiguous immunogenicity and HLA 4-digit information is complete. The classification model was built using 307 immunogenic peptides (Table 8) and 167 non-immunogenic peptides (Table 9). These peptides bind HLA-A02:01.

Currently the binding affinity of the peptide is considered as the main criteria to select immunogenic peptides. In general, binding affinity by standard programs such as NetMHCcons [24] with  $\leq 500$  nM is taken as cutoff to define immunogenic peptides. The distribution of binding affinity for the HLA-A\*02:01 peptides is shown in FIG. 3. If we consider  $\leq 500$  nM as cutoff to define immunogenic peptides then the sensitivity is 74.5% whereas the specificity is only 27.6%. FIG. 3B demonstrates that HLA binding does not predict immunogenic peptides because both non-immunogenic and immunogenic peptides can bind HLA with high affinity (FIG. 3B).

### Feature Construction and Selection

In order to generate features that will discriminate the TCR-binding peptides from the non-binders, we analyzed the physico-chemical composition of the amino acids and their positional biases in the 9-mer

























phobicity from chx to wat 2 3 RADA880104 hydro- Transfer free energy phobicity from chx to oct 2 3 WILM950104 hydro- Hydrophobicity phobicity coefficient in RP- HPLC 2 5,6 BULH740102 hydro- Apparent partial phobicity specific volume 2 6 CIDH920103 hydro- Normalized phobicity hydrophobicity scales for alpha + beta-proteins 2 6,7 RADA880107 hydro- Energy transfer from phobicity out to in (95% buried) 2 6,7 PONP800103 hydro- Average gain ratio in phobicity surrounding hydrophobicity 2 1,2,8,9 KANM800104 hydro- Average relative phobicity probability of inner beta-sheet 2 1,2,3,4,5, ZASB820101 hydro- Dependence of 6,7,8,9 phobicity partition coefficient on ionic strength 2 1 SUEM840102 helix/turn Zimm-Bragg parameter  $\sigma \times 1.0E4$  2 1,2 PALJ810108 helix/turn Normalized frequency of alpha- helix in alpha + beta class 2 1,2 LEVM780104 helix/turn Normalized frequency of alpha- helix 2 1,2 RICJ880104 helix/turn Relative preference value at N1 2 2 GEIM800109 helix/turn Aperiodic indices for alpha-proteins 2 2 ROBB760111 helix/turn Information measure for C-terminal turn 2 2 QIAN880112 helix/turn Weights for alpha- helix at the window position of 5 2 2,3 CHOP780212 helix/turn Frequency of the 1st residue in turn 2 2,3 BUNA790101 helix/turn alpha-NH chemical shifts 2 2,3 RICJ880114 helix/turn Relative preference value at C1 2 3 RACS820103 helix/turn Average relative fractional occurrence in AL 2 3,4 RICJ880109 helix/turn Relative preference value at Mid 2 4,5 RICJ880113 helix/turn Relative preference value at C2 2 5,6 RACS820105 helix/turn Average relative fractional occurrence in E0 2 6 CHOP780213 helix/turn Frequency of the 2nd residue in turn 2 6 RACS820106 helix/turn Average relative fractional occurrence in ER 2 6 PALJ810107 helix/turn Normalized frequency of alpha- helix in all-alpha class 2 6 QIAN880106 helix/turn Weights for alpha- helix at the window position of -1 2 6,7 MAXF760103 helix/turn Normalized frequency of zeta R 2 6,7 QIAN880137 helix/turn Weights for coil at the window position of 4 2 7,8 QIAN880101 helix/turn Weights for alpha- helix at the window position of -6 2 8,9 QIAN880102 helix/turn Weights for alpha- helix at the window position of -5 2 8,9 NAKH920101 helix/turn AA composition of CYT of single- spanning proteins 2 3,4,5,6, RICJ880109 helix/turn Relative preference 7,8 value at Mid .sup.1 Amino acid index .sup.2 PepLib library ID

#### Example 1a

A method of selecting immunogenic peptide from a peptide sequence TCR binding prediction Features of amino acids at each of the 9 positions of the 9-mer peptide considered for predicting immunogenicity

TABLE-US-00012 Feature number Feature value Feature ID Feature description f1 Average value of RICJ880105.sup.1 Relative preference value position 5, 6 at N2 (Richardson- Richardson) f2 Average value of QIAN880107.sup.1 Weights for alpha-helix at position 1, 2, 8, 9 the window position of 0 (Qian-Sejnowski) f3 Average value of YUTK870103.sup.1 Activation Gibbs energy position 8, 9 of unfolding f4 Value of position FNSA.2.sup.2 a combination of surface 3 area and partial charge f5 Average value of VASM830101.sup.1 Relative population of position 6, 7 conformational state A (Vasquez et al.) f6 Average value of ROBB760108.sup.1 Information measure for position 6, 7 turn (Robson-Suzuki) f7 Average value of NAKH920106.sup.1 AA composition of CYT of position 1-9 multi-spanning proteins (Nakashima-Nishikawa) f8 Average value of QIAN880139.sup.1 Weights for coil at the position 2, 3 window position of 6 (Qian- Sejnowski) f9 Average value of QIAN880138.sup.1 Weights for coil at the position 7, 8 window position of 5 (Qian- Sejnowski) f10 Average value of CHAM830103.sup.1 The number of atoms in the position 1-9 side chain labelled 1 + 1 (Charton-Charton) f11 Average value of YUTK870103.sup.1 Activation Gibbs energy of position 5, 6 unfolding f12 Average value of MITS020101.sup.1 Amphiphilicity index position 1, 2 (Mitaku et al.) f13 Value of position PNSA.1.AUTO.sup.2 a combination of surface 2 area and partial charge f14 Value of position KARS160118.sup.1 Average weighted atomic 3 number or degree based on atomic number in the graph (Karkbara-Knisley) f15 Average value of YUTK870104.sup.1 Activation Gibbs energy of position 8, 9 unfolding

Rules for predicting immunogenicity based on the features of amino acids at each of the 9 positions of the 9-mer peptide. The rules specify the range of parameters that define the identity of each amino acid at each position of the 9-mer peptide

Rule 1:  $f1 \leq 0.5$

Rule 2:  $f1 > 0.5$  AND  $f2 \leq -0.77$

Rule 3:  $f1 > 0.5$  AND  $f2 > -0.77$  AND  $f3 \leq 17.75$

Rule 4:  $f1 > 0.5$  AND  $f2 > -0.77$  AND  $f3 > 17.75$  AND  $f4 \leq -0.34$  AND  $f5 \leq 0.2055$

Rule 5:  $f1 > 0.5$  AND  $f2 > -0.77$  AND  $f3 > 17.75$  AND  $f4 > -0.34$  AND  $f6 \leq -5.5$

Rule 6:  $f1 > 0.5$  AND  $f2 > -0.77$  AND  $f3 > 17.75$  AND  $f4 > -0.34$  AND  $f6 > -5.5$  AND  $f7 \leq 45.56$  AND  $f8 > -0.055$

Rule 7:  $f1 > 0.65$  AND  $f2 > -0.77$  AND  $f3 > 17.75$  AND  $f4 > -0.34$  AND  $f6 > -5.5$  AND  $f7 > 45.56$  AND  $f8 > -0.055$  AND  $f9 \leq -0.23$  AND  $f10 > 7.0$

Rule 8:  $f1 > 0.5$  AND  $f2 > -0.77$  AND  $f3 > 17.75$  AND  $f4 > -0.34$  AND  $f6 > -5.5$  AND  $f7 > 45.56$  AND  $f9 > -0.23$  AND  $f12 \leq 0.625$  AND  $f13 \leq 0.144401$  AND  $f13 > -0.303435$  AND  $f14 \leq 6.8$  AND  $f15 \leq 18.04$

Rule 9:  $f1 > 0.5$  AND  $f2 > -0.77$  AND  $f3 > 17.75$  AND  $f4 > -0.34$  AND  $f6 > -5.5$  AND  $f7 > 45.56$  AND  $f9 > -0.23$  AND  $f12 \leq 0.625$  AND  $f13 \leq 0.144401$  AND  $f14 > 6.8$  AND  $f11 \leq 17.92$

Rule 10:  $f1 > 0.5$  AND  $f2 > -0.77$  AND  $f3 > 17.75$  AND  $f4 > -0.34$  AND  $f6 > -5.5$  AND  $f7 > 45.56$  AND  $f9 > -0.23$  AND  $f12 \leq 0.625$  AND  $f13 > 0.144401$

### Rules for Rank Ordering of Immunogenic Peptides

TABLE-US-00013 TABLE 12 Method of rank ordering immunogenic peptides Steps as shown in FIG. 1  
 Output from the steps Score TCR binding (Step-10) Positive by Ensemble model-2 and 3 3 Positive by Ensemble model 3 only 2 Positive by Ensemble model-2 only 1 Negative by both Ensemble 0 model 2 and 3  
 MHC binding (IC.sub.50)  $\leq 100$  nM 4 (Step-11)  $> 100$  nM,  $\leq 500$  nM 3  $> 500$  nM,  $\leq 1000$  nM 2  $> 1000$  nM  
 1 Expression of the =0 0 mutant allele 1-5 (read count) I (Step-7) 6-10 (read count) 2 11-50 (read count) 3  
 $> 50$  (read count) 4 TAP binding (Step-12)  $< 0.5$  3  $\geq 0.5$  1 Proteasomal cleavage  $< 10.0$  1 (Step-13)  $\geq 10$  3  
 Scores are combined to create a rank ordered score for each peptide.

### Example 2

The example demonstrates an exemplary methodology for predicting immunogenic peptide from a human Head and Neck cancer sample starting from human cancer tissue sample

### Exome Sequencing

The exome sequencing was performed for the tumor and normal samples. The exome capturing was performed using Agilent SureSelect Human All Exon V5 kit. The RNA sequencing (RNA-seq) was performed for the total RNA extracted after Ribo-depletion of tumor sample RNA. All paired-end sequencing was performed using Illumina HiSeq 2500 platform. Total data obtained for the exome-seq and RNA-seq sample exceeds 12 Gb and more than 90% of data exceed Q30 (shown in Table 12).

The exome-seq data is first pre-processed, where we remove the low quality reads/bases and adapter sequences. The pre-processed reads is then aligned to the human reference genome (hg19) using BWA program with default parameters. Then, we apply GATK-best practices where we remove the duplicate reads using Picard tools and re-align, re-calibrate using GATK and keep the file ready for somatic mutation identification (Table 13). The somatic mutations in the samples are identified using Strelka program. After this, only the quality passed and on-target mutations are processed further. A total of 222 mutations were identified in this sample. Of these 210 are SNPs and 12 are Indels (Table 14). Of the total coding mutations, 106 of them are of missense type (Table 16).

### RNA Sequencing

The RNA-seq data is first pre-processed, where we remove the low quality reads/bases, adapter sequences and unwanted sequences like ribosomal RNA, tRNAs, repeat sequences. The pre-processed reads is then aligned to human reference transcriptome and genome using STAR aligner (Table 17). The expression of the gene is then identified using Cufflinks program.

### HLA-Typing

The RNA-seq data is then used for HLA typing [27, 28]. We used Seq2HLA program for HLA typing from RNA-seq. The Class-I HLA alleles identified for this sample is provided in Table 18. The expression of the HLA genes is provided in Table 19. The read depth of the mutant allele in RNA-seq is then calculated. Of the total mutations, we found 62 mutations with read support  $\geq 1$  in RNA-seq. These mutations are also termed as expressed mutations. The 62 mutations generated 578 unique 9-mer peptides.

### Immunogenic Peptide Identification

The peptides derived from the expressed mutations were scored for TCR-binding followed by HLA binding prediction, then TAP prediction and finally proteasomal processing. The immunogenic peptides were further ranked based on the expression level of genes and variants, affinity of HLA binding, sensitivity to proteasomal processing and binding to the transporter. We applied the ranking method to 220 unique immunogenic peptides from this Head and Neck cancer sample. The ranked peptide along with HLA information is provided in Table 20.

TABLE-US-00014 TABLE 13 Summary of data generated from head and neck cancer tumor and paired normal sample Exome-seq RNA-seq Data Metrics Blood Tumor Tumor Total reads 12,65,08,302 12,38,71,688 136,893,000 Total data (Gb) 12.65 12.39 13.69 Average read length (bp) 100 100 100 GC (%) 48.98 49.85 54.55 Average base quality (Phred) 39.90 39.74 34.97 Total data  $\geq$  Q30 (%) 96.91 96.39 90.62

TABLE-US-00015 TABLE 14 Preprocessing, alignment and coverage summary of exome sequencing data Data and analysis metrics Blood Tumor Total reads after pre-processing 12,64,41,480 12,38,71,678 Total data after pre-processing (Gb) 12.63 12.38 Average read length (bp) 99.91 99.94 Average base quality (Phred) 39.72 39.56 Data  $\leq$ Q30 (%) after pre-processing 96.96 96.45 Total aligned reads 126,390,638 123,793,462 Alignment (%) 99.96 99.94 Duplicate (%) 14.98 16.20 Panel length 5,03,90,601 5,03,90,601 Panel Coverage (%) 99.85 99.84 Panel Ontarget Region Avg. Depth 111.01 130.42 On-target (%) 62.61 75.75

TABLE-US-00016 TABLE 15 Summary of variants detected in the sample Total variants 222 Total SNPs 210 Total Indels 12 Transition SNPs 136 Transversion SNPs 74 Ts/Tv 1.84

TABLE-US-00017 TABLE 16 Classification of protein-altering variants Variant Class # of mutations Missense 106 Frameshift 3 InFrame 3 Total 112 Missense - Genetic alteration that results in a different amino acid. Frameshift - Genetic alteration that changes the reading frame. This typically results in a string of different amino acids substitutions before encountering a stop codon. InFrame - Genetic alteration that results in either deletion or insertion of one or more amino acids.

TABLE-US-00018 TABLE 17 Pre-processing and alignment summary of RNA sequence data Read Count After Adapter Trimming 133,225,190 Read Count After Contamination Removal 92,623,074 Reads Aligned 75,489,728 Reads Unaligned 17,133,346 Reads Aligned % 81.50 % data lost after Pre-Processing 32.34

TABLE-US-00019 TABLE 18 HLA class I alleles present in the sample HLA-A HLA-A33:03, HLA-A02:01 HLA-B HLA-B58:01, HLA-B35:01 HLA-C HLA-C03:02, HLA-C04:01

TABLE-US-00020 TABLE 19 Expression of HLA class I genes in the sample HLA gene Gene Expression (RPKM) HLA-A 657.30 HLA-B 987.41 HLA-C 691.26

TABLE-US-00021 TABLE 20 Rank ordered list of immunogenic peptides from the mutations in head and neck cancer sample Amino Mutant acid Peptide Rank Gene change (9mer) HLA Types 1 PIK3CA p.E542K strdpls(K)i HLA-B35:01,HLA- (SEQ ID A02:01,HLA-B58:01,HLA- NO.: 513) C04:01,HLA-C03:02,HLA-A33:03 2 BRPF3 p.R570W rllieli(W)k HLA-B35:01,HLA- (SEQ ID A02:01,HLA-858:01,HLA- NO.: 514) C04:01,HLA-C03:02,HLA- A33:03 3 ZBTB6 p.E196Q stvesIts(Q) HLA-B35:01,HLA- (SEQ ID A02:01,HLA-B58:01,HLA- NO.: 515) C04:01,HLA-C03:02,HLA- A33:03 3 BRPF3 p.R570W llieli(W)kr HLA-A33:03 (SEQ ID NO.: 516) 5 BRPF3 p.R570W lieli(W)kre HLA-B35:01,HLA- (SEQ ID A02:01,HLA-B58:01,HLA- NO.: 517) C04:01,HLA-C03:02,HLA- A33:03 6 PIK3CA p.E542K (K)iteigekdf HLA-B35:01,HLA- (SEQ ID A02:01,HLA-B58:01,HLA- NO.: 518) C04:01,HLA-C03:02,HLA- A33:03 7 ZBTB6 p.E196Q lts(Q)rkemk HLA-B35:01,HLA- (SEQ ID A02:01,HLA-B58:01,HLA- NO.: 519)





5349PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 34Thr Val Gly Gln Arg Ile Gly Ser Val1  
5359PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 35Arg Thr Pro Glu Val Gln Gly Arg Val1  
5369PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 36Arg Ser Leu Leu Ala Cys Cys Gln Leu1  
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5389PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 38Arg Trp Leu Leu Val Ser Ser Pro Pro1  
5399PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 39Phe Trp Arg Ser Leu Leu Ala Cys Cys1  
5409PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 40Val Val Val Val Phe Ala Val Cys Trp1  
5419PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 41Thr Cys Asn Ser Arg Gln Ala Ala Leu1  
5429PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 42Pro Val Gln Arg Leu Pro Phe Ser Thr1  
5439PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 43Ala Leu Ser Arg Pro Gly Leu Leu Arg1  
5449PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 44Glu Pro Ile Tyr Met Tyr Ser Thr Met1  
5459PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 45Val Val Gly Arg Ser Val Ala Ile Gly1  
5469PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 46His Gly Arg Ala Asp Leu Ile Arg Leu1  
5479PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 47Ser Gly Val Gly Lys Ser Ala Leu Thr1  
5489PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 48Arg Tyr Pro Val Gln Arg Leu Pro Phe1  
5499PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 49Asp Leu Ile Arg Leu Leu Leu Lys His1  
5509PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 50Ala Asp Leu Ile Arg Leu Leu Leu Lys1  
5519PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 51Leu His Ser Leu Trp Thr Cys Asp Cys1  
5529PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 52Val Ala Ile Gly Pro Arg Glu Gln Trp1  
5539PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 53Leu Ile Arg Leu Leu Leu Lys His Gly1  
5549PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 54Ser Ala Thr Val Thr Ala Phe Trp Arg1  
5559PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 55Gly Ser Val Ser Phe Gly Thr Val Tyr1  
5569PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 56Val Gln Gly Arg Val Pro Thr Leu Glu1  
5579PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 57Pro Gln Ala Arg Ala Val His Leu Pro1  
5589PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 58Leu Ser Arg Pro Gly Leu Leu Arg Gln1  
5599PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 59Leu Arg Glu Ala Ser Pro Trp Val Arg1  
5609PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 60Arg Pro Glu Val Arg Lys Thr Ala Ser1  
5619PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 61Leu His Gly Arg Ala Asp Leu Ile Arg1  
5629PRTHomo sapiensPEPTIDE(1)..(9)selected peptide variant with a predicted ability to interact with the TCR and may or can serve as a mammalian tumor vaccine 62Gln Gly Arg Val Pro Thr Leu Glu Arg1













binding algorithm 284Leu Val Leu Ile Leu Tyr Leu Cys Val1 52859PRTHomo sapiensPEPTIDE(1)..  
(9)Immunogenic peptide used for developing the TCR-binding algorithm 285Gly Met Ser Arg Ile Gly Met  
Glu Val1 52869PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 286Phe Leu Ser His Asp Phe Thr Leu Val1 52879PRTHomo sapiensPEPTIDE(1)..  
(9)Immunogenic peptide used for developing the TCR-binding algorithm 287Cys Ile Asn Gly Val Cys Trp  
Ser Val1 52889PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 288Ser Ile Thr Glu Val Glu Cys Phe Leu1 52899PRTHomo sapiensPEPTIDE(1)..  
(9)Immunogenic peptide used for developing the TCR-binding algorithm 289Arg Leu Glu Arg Lys Trp Leu  
Asp Val1 52909PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 290Ser Ile Asp Gln Leu Cys Lys Thr Phe1 52919PRTHomo sapiensPEPTIDE(1)..  
(9)Immunogenic peptide used for developing the TCR-binding algorithm 291Gln Leu Phe Asn His Thr Met  
Phe Ile1 52929PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 292Met Ile Met Gln Gly Gly Phe Ser Val1 52939PRTHomo sapiensPEPTIDE(1)..  
(9)Immunogenic peptide used for developing the TCR-binding algorithm 293Lys Cys Ile Asp Phe Tyr Ser  
Arg Ile1 52949PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 294Ser Leu Lys Lys Asn Ser Arg Ser Leu1 52959PRTHomo sapiensPEPTIDE(1)..  
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Thr Thr1 52969PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 296Met Thr Ile Ile Phe Leu Ile Leu Met1 52979PRTHomo sapiensPEPTIDE(1)..  
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binding algorithm 298Tyr Leu Glu Pro Gly Pro Val Thr Ala1

52999PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding  
algorithm 299Ile Leu Asp Lys Lys Val Glu Lys Val1 53009PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic  
peptide used for developing the TCR-binding algorithm 300Leu Ala Leu Leu Leu Leu Asp Arg Leu1  
53019PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding  
algorithm 301Phe Ile Asp Lys Phe Thr Pro Pro Val1 53029PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic  
peptide used for developing the TCR-binding algorithm 302Gly Ile Leu Glu Phe Val Phe Thr Leu1  
53039PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding  
algorithm 303Tyr Leu Val Ser Ile Phe Leu His Leu1 53049PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic  
peptide used for developing the TCR-binding algorithm 304Phe Val Val Pro Ile Leu Leu Lys Ala1  
53059PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding  
algorithm 305Cys Leu Pro Ala Cys Val Tyr Gly Leu1 53069PRTHomo sapiensPEPTIDE(1)..  
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Val Thr1 53079PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 307Thr Leu Leu Asp His Ile Arg Thr Ala1 53089PRTHomo sapiensPEPTIDE(1)..  
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Glu Val1 53099PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 309Ala Val Ala Asp His Val Ala Ala Val1 53109PRTHomo sapiensPEPTIDE(1)..  
(9)Immunogenic peptide used for developing the TCR-binding algorithm 310Thr Leu Asn Asp Leu Glu Thr  
Asp Val1 53119PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 311Thr Leu Leu Ala Asn Val Thr Ala Val1 53129PRTHomo sapiensPEPTIDE(1)..  
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Arg Leu1 53139PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 313Ala Leu Pro His Ile Ile Asp Glu Val1 53149PRTHomo sapiensPEPTIDE(1)..  
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Ser Leu1 53159PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 315Thr Leu Thr Ser Tyr Trp Arg Arg Val1 53169PRTHomo sapiensPEPTIDE(1)..  
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Glu Leu1 53179PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 317Ser Leu Met Asp Leu Leu Ser Ser Leu1 53189PRTHomo sapiensPEPTIDE(1)..  
(9)Immunogenic peptide used for developing the TCR-binding algorithm 318Phe Leu Thr Ser Val Ile Asn  
Arg Val1 53199PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 319Gly Ile Leu Asp Phe Gly Val Lys Leu1 53209PRTHomo sapiensPEPTIDE(1)..  
(9)Immunogenic peptide used for developing the TCR-binding algorithm 320Gln Leu Val Gln Ser Gly Ala  
Glu Val1 53219PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
binding algorithm 321Tyr Leu Leu Lys Pro Val Gln Arg Ile1 53229PRTHomo sapiensPEPTIDE(1)..





binding algorithm 399Arg Val Asn Arg Leu Ile Ile Trp Val1 54009PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the TCR-binding algorithm 400Ser Leu Met Ser Gly Val Glu  
 Pro Leu1 54019PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 401Thr Leu Asp Tyr Lys Pro Leu Ser Val1 54029PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the TCR-binding algorithm 402Ser Leu Phe Asn Thr Val Ala  
 Thr Leu1 54039PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 403Val Leu Leu Arg His Ser Lys Asn Val1 54049PRTHomo sapiensPEPTIDE(1)..  
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 Leu Ile1 54059PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 405Ser Leu Leu Met Trp Ile Thr Gln Cys1 54069PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the

TCR-binding algorithm 406Gly Leu Asn Asp Tyr Leu His Ser Val1 54079PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the TCR-binding algorithm 407Ala Met Ala Ser Thr Glu Gly  
 Asn Val1 54089PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 408Gly Leu Arg Glu Asp Leu Leu Ser Leu1 54099PRTHomo sapiensPEPTIDE(1)..  
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 Cys Val1 54109PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 410Ala Leu Ala Ile Ile Ala Val Leu1 54119PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the TCR-binding algorithm 411Leu Gln Leu Pro Gln Gly Thr  
 Thr Leu1 54129PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 412Phe Leu Trp Glu Asp Gln Thr Leu Leu1 54139PRTHomo sapiensPEPTIDE(1)..  
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 Leu Leu1 54149PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 414Gly Ile Trp Gly Phe Val Phe Thr Leu1 54159PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the TCR-binding algorithm 415Phe Ala Asn His Lys Phe Thr  
 Leu Val1 54169PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 416Gln Met Met Arg Asn Glu Phe Arg Val1 54179PRTHomo sapiensPEPTIDE(1)..  
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 Leu Leu1 54189PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 418Gly Ile Leu Thr Val Ser Val Ala Val1 54199PRTHomo sapiensPEPTIDE(1)..  
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 Ile Val1 54209PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 420Ala Leu Tyr Asp Val Val Ser Lys Leu1 54219PRTHomo sapiensPEPTIDE(1)..  
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 binding algorithm 422Leu Leu Gly Arg Asn Ser Phe Glu Val1 54239PRTHomo sapiensPEPTIDE(1)..  
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 Pro Ile1 54249PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 424Met Leu Trp Gly Tyr Leu Gln Tyr Val1 54259PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the TCR-binding algorithm 425Asn Leu Leu Thr Thr Pro Lys  
 Phe Thr1 54269PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 426Thr Leu Tyr Ala Val Ala Thr Thr Ile1 54279PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the TCR-binding algorithm 427Phe Leu Lys Gln Gln Tyr Met  
 Asn Leu1 54289PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 428Lys Asp Leu Val Leu Leu Ala Thr Ile1 54299PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the TCR-binding algorithm 429Leu Leu Val Ser Glu Ile Asp  
 Trp Leu1 54309PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 430Lys Leu Asn Pro Met Leu Ala Lys Ala1 54319PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the TCR-binding algorithm 431Val Ile Phe Asp Phe Leu His  
 Cys Ile1 54329PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 432Phe Ala Asn Asn Glu Phe Thr Leu Val1 54339PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the TCR-binding algorithm 433Val Leu Cys Leu Arg Pro Val  
 Gly Ala1 54349PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 434Ser Leu Phe Leu Gly Ile Leu Ser Val1 54359PRTHomo sapiensPEPTIDE(1)..  
 (9)Immunogenic peptide used for developing the TCR-binding algorithm 435Ala Leu Ala His Gly Val Arg  
 Ala Leu1 54369PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-  
 binding algorithm 436Ala Leu Leu Ala Leu Thr Arg Ala Ile1 54379PRTHomo sapiensPEPTIDE(1)..



(9)Immunogenic peptide used for developing the TCR-binding algorithm 476Ala Leu Leu Glu Asp Pro Val Gly Thr1 54779PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 477Phe Val Asn Tyr Asn Phe Thr Leu Val1 54789PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 478Trp Gln Trp Glu His Ile Pro Pro Ala1 54799PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 479Val Met Leu Phe Ile Leu Ala Gly Leu1 54809PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 480Met Thr Tyr Ala Ala Pro Leu Phe Val1 54819PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 481Tyr Leu Lys Lys Ile Lys Asn Ser Leu1 54829PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 482Ala Met Ala Gly Ala Ser Thr Ser Ala1 54839PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 483Asn Met Leu Ser Thr Val Leu Gly Val1 54849PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 484Ile Leu Ala Lys Phe Leu His Trp Leu1 54859PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 485Lys Leu Gly Pro Gly Glu Glu Gln Val1 54869PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 486Ser Val Tyr Asp Phe Phe Val Trp Leu1 54879PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 487Val Leu Thr Ser Glu Ser Met His Val1 54889PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 488Ser Leu Ser Arg Phe Ser Trp Gly Ala1 54899PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 489Arg Met Leu Gly Asp Val Met Ala Val1 54909PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 490Tyr Ile Leu Glu Glu Thr Ser Val Met1 54919PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 491Ile Leu Asp Ala His Ser Leu Tyr Leu1 54929PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 492Gly Ile Phe Glu Asp Arg Ala Pro Val1 54939PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 493Thr Val Cys Gly Gly Ile Met Phe Leu1 54949PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 494Gly Leu Cys Pro His Cys Ile Asn Val1 54959PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 495Ala Phe Leu Gly Glu Arg Val Thr Leu1 54969PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 496Asn Gly Val Arg Val Leu Ala Thr Ala1 54979PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 497Gln Leu Leu Asn Ser Val Leu Thr Leu1 54989PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 498Ile Leu His Thr Asn Met Pro Asn Val1 54999PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 499Ala Ile Thr Glu Val Glu Cys Phe Leu1 55009PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 500Gly Met Asp Pro Arg Met Cys Ser Leu1 55019PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 501Ala Ile Leu Ile Arg Val Arg Asn Ala1 55029PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 502Lys Thr Val Leu Glu Leu Thr Glu Val1 55039PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 503Val Leu His Lys Arg Thr Leu Gly Leu1 55049PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 504Met Gly Asn Gly Cys Leu Arg Ile Val1 55059PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 505Leu Val Met Ala Gln Leu Leu Arg Ile1 55069PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 506Ala Met Leu Asp Leu Leu Lys Ser Val1 55079PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 507Ile Ala Asp Ala Ala Leu Ala Ala Leu1 55089PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 508Asp Leu Ser Leu Arg Arg Phe Met Val1 55099PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 509Leu Gln Asp Ile Glu Ile Thr Cys Val1 55109PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 510Lys Leu Gln Glu Gln Gln Ser Asp Leu1 55119PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 511Phe Leu Thr Cys Thr Asp Arg Ser Val1 55129PRTHomo sapiensPEPTIDE(1)..(9)Immunogenic peptide used for developing the TCR-binding algorithm 512Ser Val Gly Gly Val Phe Thr Ser Val1 55139PRTHomo sapiensPEPTIDE(1)..(9)immunogenic peptides from the mutations in head and neck cancer 513Ser Thr Arg Asp Pro Leu Ser Lys Ile1 55149PRTHomo sapiensPEPTIDE(1)..(9)immunogenic peptides from the

mutations in head and neck cancer 514Arg Leu Leu Ile Glu Leu Ile Trp Lys1 55159PRTHomo sapiensPEPTIDE(1)..(9)immunogenic peptides from the mutations in head and neck cancer 515Ser Thr Val Glu Ser Leu Thr Ser Gln1 55169PRTHomo sapiensPEPTIDE(1)..(9)immunogenic peptides from the mutations in head and neck cancer 516Leu Leu Ile Glu Leu Ile Trp Lys Arg1 55179PRTHomo sapiensPEPTIDE(1)..(9)immunogenic peptides from the mutations in head and neck cancer 517Leu Ile Glu Leu Ile Trp Lys Arg Glu1 55189PRTHomo sapiensPEPTIDE(1)..(9r)immunogenic peptides from the mutations in head and neck cancer 518Lys Ile Thr Glu Gln Glu Lys Asp Phe1 55199PRTHomo sapiensPEPTIDE(1)..(9)immunogenic peptides from the mutations in head and neck cancer 519Leu Thr Ser Gln Arg Lys Glu Met Lys1 5

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